

Editorial Team

Editors-in-Chief:

Fikret Çelebi, PhD, Professor of Veterinary Physiology; Atatürk University, **TURKEY**

Daryoush Babazadeh ([ORCID ID](#); [Publons](#); [Fulle Member of WAME](#); [Member of IAVE](#); Email: daryoush.babazadeh@shirazu.ac.ir); DVM, DVSc, PhD of Avian/Poultry Diseases, School of Veterinary Medicine, Shiraz University, Shiraz, **IRAN**

Managing Editor:

Alireza Sadeghi, DVM, Faculty of Veterinary medicine, Tabriz Branch, Islamic Azad University, Tabriz, **IRAN**; Email: alirezavet86@gmail.com

Associate Editors

Anjum Sherasiya, Ex-Veterinary Officer, Star, Gulshan Park, NH-8A, Chandrapur Road, Wankaner - 363621, Dist. Morbi (Gujarat), **INDIA**

Arman Moshaveri, DVM, Faculty of Veterinary Medicine, Karaj Branch, Islamic Azad University, Karaj, **IRAN**

Ashraf Fathy Said Awad, PhD, Genetics and Genetic Engineering, Animal, Wealth Development Department, Faculty of Veterinary, Medicine, Zagazig University, **EGYPT**

Konstantinos Koutoulis, DVM, PhD; Avian Pathology; Faculty of Veterinary Science, University of Thessaly, Terma Trikalon 224, 43100 Karditsa, **GREECE**

Mahendra Pal, PhD. Ex-Professor of Veterinary Public Health, Department of Microbiology, Immunology and Public Health, College of Veterinary Medicine, Addis Ababa University, **ETHIOPIA**

Saeid Chekani Azar, PhD, Animal Physiology; Faculty of Veterinary Medicine, Atatürk University, Erzurum, **TURKEY**; [ORCID ID](#), [Google Scholar](#)

Thakur Krishna Shankar Rao, PhD, Assistant professor, Vanabandhu College of Veterinary Science & Animal Husbandry, Navsari Agricultural University, Navsari Gujarat, **INDIA**

Thandavan Arthanari Kannan, PhD, Full professor, Centre for Stem Cell Research and Regenerative Medicine Madras Veterinary College Tamil Nadu Veterinary and Animal Sciences university Chennai-600007, **INDIA**

Tohid Vahdatpour, PhD, Assistant Prof., Physiology; Dep. Animal Sciences, Shabestar Branch, Islamic Azad University, Shabestar, **IRAN**

Wesley Lyeverton Correia Ribeiro, MSc, DVM, Animal Health, Veterinary Parasitology, and Public Health, Animal welfare; College of Veterinary Medicine, State University of Ceará, Av. Paranjana, 1700, Fortaleza, **BRAZIL**

Language Editor:

Ali Fazel, Master of arts in T.E.S.O.L. University of Nottingham, Semenyih, Selanger, **MALAYSIA**

Faezeh Modarresi-Ghazani, Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, **IRAN**

Reviewers

Ahmed Mohamed Ammar, Professor of Microbiology, Faculty of Veterinary Medicine, Zagazig University, **EGYPT**

Alireza Koochakzadeh, DVM, PhD of Bacteriology, Faculty of Veterinary Medicine, University of Tehran, Tehran, **IRAN**

AKM Mostafa Anower, PhD, Patuakhali Science and Technology University, Department of Microbiology and Public Health, Faculty of Animal Science and Veterinary Medicine, **BANGLADESH**

Ali Halajian, PhD, DVM, Parasitology; Dep. Biodiversity, School of Molecular & Life Sciences, Faculty of Science and Agriculture, University of Limpopo, **SOUTH AFRICA**

Ghader Najafi, PhD in Animal Physiology, Ankara University, Ankara, **TURKEY**; Assistant Prof. in Faculty of Veterinary Medicine, Islamic Azad University, Urmia Branch, Urmia, **IRAN**

Hazim Jabbar Al-Daraji, PhD, Professor of Avian Reproduction and Physiology; University of Baghdad, College of Agriculture, Abu-Ghraib, Baghdad, **IRAQ**

KARAMALA SUJATHA, MVSc, PhD, Associate Professor, Department of Veterinary Pathology, College of Veterinary Science, Sri Venkateswara Veterinary University, Tirupati – 517502, Andhra Pradesh, **INDIA**

Khalid Mohammed Elamin Osman, PhD, Associate Professor of Animal Production; University of Gezira, Faculty of Animal Production, **SUDAN**

Kuastros Mekonnen Belaynehe, Seoul National University, South Korea/ National Animal Health diagnostics and Investigation Center, **ETHIOPIA**

Mahdi Alyari Gavaher, DVM, DVSc, Faculty of Veterinary Medicine, Karaj Branch, Islamic Azad University, Karaj, **IRAN**

Manish Kumar, Prof. Dr. Pharmacology, Ethnomedicine, Society of Education (SOE), **INDIA**

Mojtaba Mohseni, DVM, DVSc (PhD) Student of Large Animal Internal Medicine, Faculty of Veterinary Medicine, Urmia University, Urmia, **IRAN**

Muhammad Abdullahi Mahmud, DVM, MSc, SENIOR LECTURER, Department of Animal Health and Production Technology, Niger State College of Agriculture, Mokwa, Niger State, **NIGERIA**

Muhammad Moin Ansari, BVSc & AH, MVSc, PhD (IVRI), NET (ICAR), Dip.MLT, CertAW, LMIVA, LMISVS, LMISVM, MHM, Sher-e-Kashmir University of Agricultural Sciences and Technology of Kashmir, Faculty of Veterinary Sciences and Animal Husbandry, Division of Veterinary Surgery and Radiology, Shuhama, Alastang, Srinagar-190006 Jammu & Kashmir, **INDIA**

Muhammad Saeed, PhD (Student), Animal Nutrition and Feed Science, College of Animal Sciences and Feed technology, Northwest A&F University, Yangling, 712100, **CHINA**

Nunna Veera Venkata Hari Krishna, PhD, Assistant Professor, Dept. of Veterinary Surgery and Radiology NTR College of Veterinary Science, Gannavaram – 521 102, A.P., **INDIA**

Osman Erganiş, PhD. Professor of Microbiology; Department of Microbiology, Faculty of Veterinary Medicine, Selcuk University, Konya, **TURKEY**

Rafiqul Islam, Animal Scientist, Krishi Vigyan Kendra, Dhubri, Assam Agricultural University, Bilasipara, PO: Bilasipara, District: Dhubri, State: Assam, **INDIA**

Shewangzaw Addisu Mekuria, BSc, MSc, Instructor, department of Animal Production and Extension, University of Gondar, P. O. Box 196, Gondar, **ETHIOPIA**

Siamk Sandoughchian, PhD, Immunology; Department of Immunology, Faculty of Medical Sciences, Juntendo University, **JAPAN**

Terry Ansah, PhD., Nutrition - Ruminants; University for Development Studies-Ghana and Harper Adams University College, **UNITED KINGDOM**

Tesfaheywet Zeryehun Shiferaw, DVM, MSc, Associate Professor, College of Veterinary Medicine Haramaya University, P.O.Box-301, Dire Dawa, **ETHIOPIA**

Thakur Krishna Shankar Rao, PhD, Assistant professor, Vanabandhu College of Veterinary Science & Animal Husbandry, Navsari Agricultural University, Navsari Gujarat, **INDIA**

Vassilis Papatsiros, Professor, Dietary input, Animal and Feed interactions; Porcine Medicine, Faculty of Veterinary Medicine, University of Thessaly, Trikalon str 224, GR 43100, **GREECE**

Wafaa Abd El-Ghany Abd El-Ghany, PhD, Assistant Prof. of Poultry and Rabbit Diseases; Poultry and Rabbit Diseases Department, Faculty of Veterinary Medicine, Cairo University, Giza, **EGYPT**

Varij Nayan, BVSc, MVSc, PhD Scientist (Animal Biochemistry), Animal Physiology and Reproduction Division, ICAR-Central Institute for Research on buffaloes (ICAR-CIRB), Hisar-125001 (Haryana) **INDIA**,

Yagoob Garedaghi, Assistant professor, PhD of Parasitology; Department of Veterinary Parasitology, Tabriz Branch, Islamic Azad University, Tabriz, **IRAN**

Sesotya Raka Pambuka, MSc, Sinta Prima Feedmill, Poultry and Aqua Feed Formulation, Sulaiman Rd 27A, West Jakarta, **INDONESIA**

Advisory Board

Ferdaus Mohd. Altaf Hossain, DVM, Microbiology, Immunology, Poultry Science, and Public Health; Sylhet Agricultural University, Bangladesh; not shah Jalal University of Science & Technology, **BANGLADESH**

Paola Roncada, PhD, Associate Prof., Pharmacokinetics, Residues of mycotoxins in food and in foodproducing species, Residue depletion studies; Veterinary Pharmacology and Toxicology, Faculty of Veterinary Medicine, University of Bologna, **ITALY**

Sina Vahdatpour, DVM-DVMS, Faculty of Veterinary medicine, Tabriz Branch, Islamic Azad University, Tabriz, **IRAN**

Tohid Vahdatpour, PhD, Assistant Prof., Physiology; Dep. Animal Sciences, Shabestar Branch, Islamic Azad University, Shabestar, **IRAN**

Volume 8 (3); September 25, 2018

Research Paper

The Epidemiology of Canine Parvovirus Enteritis in Dogs of Makurdi, Benue State, Nigeria.

Tion MT, Apaa TT, Saganuwan AS, Nwankwo HC, Tughgba T, Anumtyo TM, Amine AA, Nguetyo SA, Igoh, AF and Akpehe-Ishor W.

World Vet. J. 8(3): 48-54, 2018; pii:S232245681800005-8



Tion MT, Apaa TT, Saganuwan AS, Nwankwo HC, Tughgba T, Anumtyo TM, Amine AA, Nguetyo SA, Igoh, AF and Akpehe-Ishor W (2018). The Epidemiology of Canine Parvovirus Enteritis in Dogs of Makurdi, Benue State, Nigeria. *World Vet. J.* 8(3): 48-54.

ABSTRACT

Since its emergence, canine parvovirus enteritis has remained the most significant and important cause of enteritis in puppies between six weeks and six months of age. The aim of this study was to carry out a retrospective study in order to assess the effect of certain factors on the prevalence of the disease in diagnosed cases presented to the veterinary teaching hospital Annex, university of agriculture Makurdi from 2010 to 2016. The overall prevalence of the diagnosed cases of canine parvoviral enteritis has been at 5.7% for the past seven years. Age, sex, breed, vaccination status was showed to have been associated with the infection. Puppies of up to five months of age, males, breeds such as the Nigerian local breed, Alsatian, Caucasian, Rottweiler and Russian shepherd dogs and unvaccinated dogs are prone to this disease. Furthermore, this study revealed that early presentation and an aggressive support management of these cases had brought about a high recovery outcome of 85.4%. The distribution of the disease according to the season showed the highest incidence of cases occurred in 2012 (25.8%) and had descended down to (9%) by 2016 with most cases witnessed around January (22.5%) in dry season and June (15.7%) in wet season. The need for educating both veterinarians and dog owners about core vaccinations in dogs is paramount.

Keywords: Canine parvoviral enteritis, Dogs, Makurdi, Prevalence

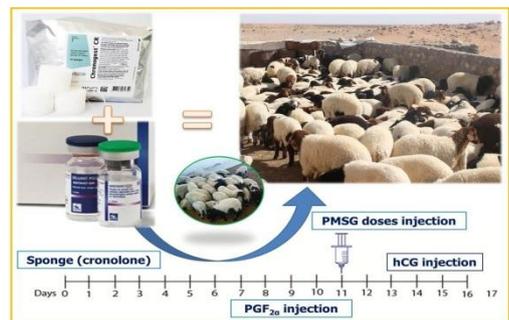
[Full text-[PDF](#)] [[MOBI](#)] [[ePUB](#)] [[AZW3](#)] [[XML](#)]

Research Paper

Application of Different Hormonal Protocols for Improving Reproductive Performance of Barki Ewes.

Ashour G, El-Bassiony MF, Dessouki ShM and El-Wakeel MA.

World Vet. J. 8(3): 55-64, 2018; pii:S232245681800006-8



Ashour G, El-Bassiony MF, Dessouki ShM and El-Wakeel MA (2018). Application of Different Hormonal Protocols for Improving Reproductive Performance of Barki Ewes. *World Vet. J.* 8(3): 55-64.

ABSTRACT

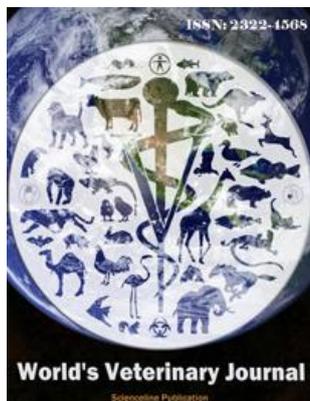
This investigation was designed to assess reproductive improvement in Barki ewes using different hormonal protocols. Seventy-five non-pregnant and non-lactating Barki ewes were randomly assigned into three equal groups (25 ewes each), namely G1, G2 and G3. (G1) served as control, while (G2) was treated with an intravaginal progestagen impregnated sponge for 12 days then was removed. On the ninth day, all treated ewes received an intramuscular injection of prostaglandin F_{2α}. (G3) was also synchronized as G2, in addition to on the 10th day ewes were received an intramuscular injection of 750 IU PMSG in descending doses for three days so that the last dose was injected at the 12th day concurrently with sponges' removal. Meanwhile, on the 14th day, all ewes in G3 were received an intramuscular injection of hCG hormone (500 IU/ewe). The results showed an increase in plasma progesterone level from the first day of pregnancy and rises up to the last day before parturition. Progesterone was found to be higher in G3 (that had higher twinning rate) than in G2 than in G1. Insignificant increase in estrus response was observed in groups (G2, G3) compared to G1 (100, 100 and 92%, respectively). Conception rate was significantly higher in G3 (100%) than G2 (92%) and G1 (88%). While, abortion rate was higher in G1 and surpassing G2 and G3 (9.0, 0.0 and 0.0 %, respectively). Lambing rate

was significantly higher in G3 as compared to G2 and G1 groups (100, 92 and 80 %, respectively). While, weaning rate was recorded to be insignificantly higher in G2 than in G1 and G3 (100.0, 95.45 and 94.29%, respectively). It could be concluded that, hormonal manipulation using intravaginal progestagen impregnated sponge and PMSG in the presence of hCG; would be a proper way for enhancing the reproductive efficiency of Barki ewes.

Keywords: Barki ewes, Synchronization, Superovulation, Twinning, Reproduction

[Full text-[PDF](#)] [[MOBI](#)] [[ePUB](#)] [[AZW3](#)] [[XML](#)]

[Archive](#)



World's Veterinary Journal

ISSN: 2322-4568

Frequency: Quarterly

Current Issue: 2018, Vol: 8, Issue: 3 (25 September)

Publisher: [SCIENCELINE](#)

World's Veterinary Journal (ISSN 2322-4568) is an international, English language, peer reviewed open access journal aims to publish the high quality material from veterinary scientists' studies

[... View full aims and scope \(www.wvj.science-line.com\)](http://www.wvj.science-line.com)

Editors-in-Chief:

Prof. Dr. Fikret Çelebi, Veterinary Physiology; Atatürk University, TURKEY;

Dr. Daryoush Babazadeh, DVM, DVSc (PhD) of Avian/Poultry Diseases, Shiraz University, Shiraz, IRAN

- WJ indexed/covered by [SCOPUS](#), [NLM Catalog \(NLM ID: 101688928\)](#), [ScopeMed](#), [RiCeST-ISC](#), [Ulrich's™/ProQuest](#), [NAAS \(Score: 3.96\)](#), [UBTIB](#), [SHERPA/RoMEO](#), [Genamic](#), [INFOBASE](#), [Index Copernicus International \(ICV 2014= 5.73\)](#) (visit [full index information](#))
- Open access full-text articles is available beginning with Volume 1, Issue 1.
- Full texts and XML articles are available in [E-Journals Database \(RiCeST\)](#).
- This journal is in full compliance with [Budapest Open Access Initiative](#) and [International Committee of Medical Journal Editors' Recommendations \(ICMJE\)](#).



[ABOUT US](#)

| [CONTACT US](#)

| [PRIVACY POLICY](#)

ScienceLine Offices:

Atatürk Univ., Erzurum 25100, Turkey, www.science-line.com

Maragheh Univ., East Azerbaijan, Iran, www.science-line.ir

Tel: +90-538 770 8824; +98-914 420 7713

Email: administrator@science-line.com



The Epidemiology of Canine Parvovirus Enteritis in Dogs of Makurdi, Benue State, Nigeria

Tion Matthew Terzungwe^{1*}, Apaa Ternenge Thaddaeus¹, Saganuwan Alhaji Saganuwan², Nwankwo Henry Chukwuebuka², Tughgba Terzungwe³, Anumtyo Theresa Mwuese⁴, Amine Aondowase Andrew³, Nguetyo Samuel Aondonenge⁴, Igoh Ann Faith⁴, Akpehe-Ishor Washima⁴

¹Department of Veterinary Medicine, College of Veterinary Medicine, University of Agriculture, Makurdi, Benue State, Nigeria.

²Department of Veterinary Physiology, Pharmacology and Biochemistry, College of Veterinary Medicine, University of Agriculture, Makurdi, Benue State, Nigeria.

³Department of Veterinary Surgery and Theriogenology, College of Veterinary Medicine, University of Agriculture, Makurdi, Benue State, Nigeria.

⁴College of Veterinary Medicine, University of Agriculture, Makurdi, Benue State, Nigeria.

*Corresponding author's Email: tions_doc@yahoo.co.uk

ABSTRACT

Since its emergence, canine parvovirus enteritis has remained the most significant and important cause of enteritis in puppies between six weeks and six months of age. The aim of this study was to carry out a retrospective study in order to assess the effect of certain factors on the prevalence of the disease in diagnosed cases presented to the veterinary teaching hospital Annex, university of agriculture Makurdi from 2010 to 2016. The overall prevalence of the diagnosed cases of canine parvoviral enteritis has been at 5.7% for the past seven years. Age, sex, breed, vaccination status was showed to have been associated with the infection. Puppies of up to five months of age, males, breeds such as the Nigerian local breed, Alsatian, Caucasian, Rottweiler and Russian shepherd dogs and unvaccinated dogs are prone to this disease. Furthermore, this study revealed that early presentation and an aggressive support management of these cases had brought about a high recovery outcome of 85.4%. The distribution of the disease according to the season showed the highest incidence of cases occurred in 2012 (25.8%) and had descended down to (9%) by 2016 with most cases witnessed around January (22.5%) in dry season and June (15.7%) in wet season. The need for educating both veterinarians and dog owners about core vaccinations in dogs is paramount.

Key words: Canine parvoviral enteritis, Dogs, Makurdi, Prevalence

INTRODUCTION

The use of dogs as companion to humans cannot be overemphasized. Dogs are used as trackers, instrument of war, and healers of both the physical and emotional problems of humans (Carmichael, 2003). The population of dogs in Nigeria is estimated to be around 4.5million (Bourn et al., 1994) and infectious diseases such as parvovirus could reduce their population. The affection humans develop towards pets continues to deepen in this twenty-first century (Daodu et al., 2017).

Canine parvovirus type 2 (CPV-2) is a highly contagious and often fatal viral disease that infects the gastrointestinal tract of dogs (Touihri et al., 2009). CPV is a small, non-enveloped with spherical capsid, consisting of three proteins and a linear, single-strand DNA virus of the family Parvoviridae (Maclachlan and Dubovi, 2010). CPV-2 infection does not have predilection for sex, age or breeds of dogs (Castro et al., 2007; Gombac et al., 2008). CPV-2 affects dogs between the ages of six weeks and six months. It seldom affects older dogs, because of their natural immunity (McCaw and Hoskins, 2006; Prittie, 2004). But some breeds such as Rottweiler, Doberman pinscher, American Pit-bull terrier, Labrador retriever and German shepherd dog are at an increased risk of infection (Smith-Carr et al., 1997; Houston et al., 1996).

Certain factors that predispose puppies to CPV-2 infection are lack of protective immunity, intestinal parasites and overcrowding, poor sanitary, and stressful environmental conditions (Smith-Carr et al., 1997; Hoskins, 1997). CPV-2 can be transmitted from infected to susceptible dogs mainly through direct transmission (faecal-oral route), or indirect transmission where dogs can also become infected through exposure to fomites such as shoes, clothing, the hands of humans, food bowls and other utensils (Carmichael, 1994; Smith-Carr et al., 1997; Hoskins, 1997; Decaro et al., 2005b; Nivy et al., 2011). CPV-2 can also be transmitted via house flies, flesh flies and blow/bottle flies (Bagshaw et al., 2014).

CPV-2 has mutated twice in the early 1970s to mid 1980s given rise to two antigenic variants namely; CPV-2a and CPV-2b (Parker et al., 2001). In 2000, a third antigenic mutant named CPV-2c emerged from Italy (Buonavoglia et al., 2001). All the three antigenic variants have been distributed in different parts of the world (Bingga et al., 2014; Touihri et al., 2009; Wilson et al., 2014).

The emergence of CPV-2 infection in Nigeria can be traced back to the mid 1980s (Ezeokoli et al., 1985). Chollom et al. (2013) discovered CPV-2 with the use of conventional polymerase chain reaction (PCR). Case reports, seroprevalence, treatment regimen, immunity following CPV vaccination and risk factors affecting the disease in Nigeria have been reported (Eghafona et al., 2007; Ezeibe Maduiké et al., 2010; Nwoha, 2011; Shima et al., 2015). CPV-2a strain has been reported in Nigeria (Dogonyaro et al., 2013; Apaa et al., 2016). The virus is often shed in the faeces of infected dogs within 4-5 days of exposure before clinical signs develop and then throughout the period of illness, and for 10 days after clinical recovery (Cynthia and Scott, 2010).

CPV-2 infection manifests as an acute haemorrhagic enteritis and myocarditis. Dogs with enteritis show depression, loss of appetite, lethargy, vomiting, high fever and severe mucoid or bloody and foul smelly diarrhea (Lamm and Rezabek, 2008; Prittie, 2004). Presently, myocarditis though seen can develop in puppies less than eight weeks old dog born to unvaccinated bitches (Hoskins, 1997).

Definitive diagnosis is done by the detection of CPV-2 in the faeces of affected dogs, serology, and necropsy and histopathology (Pollock and Carmichael, 1988). Other methods of detection include electron microscopy, viral isolation, fecal hemagglutination, latex agglutination, counter-immunoelectrophoresis, immunochromatography, PCR (Macintire and Smith-Carr, 1997; Pollock and Carmichael, 1988; Desario et al., 2005; JinSik et al., 2006) which is more sensitive and reliable than traditional techniques (Desario et al., 2005).

CPV infection can be managed by aggressive symptomatic and supportive therapy (Prittie, 2004; Brown and Otto, 2008) involving fluid therapy, antibiotic, antiemetic, nutritional support, antiviral treatments and pain management (Mylonakis et al., 2016). Prevention is by vaccination of dogs with either attenuated or modified live vaccines (Martella et al., 2005). But maternally derived antibodies protect neonates as well (10 days) and it interferes with vaccines. Thus, causing vaccine failures (Pollock and Carmichael, 1982).

In Makurdi metropolis, there have been several reported cases of morbidity and mortality of CPV in dogs but there is paucity of information on prevalence, morbidity and mortality rates of the disease. Hence epidemiology of canine parvovirus infection was studied at Veterinary Teaching Hospital (VTH) of the University of Agriculture Makurdi (UAM).

MATERIALS AND METHODS

Study area

This study was conducted in Makurdi metropolis, Benue State, Nigeria. Makurdi, the capital of Benue state lies between latitude 7° 15' - 7° 45'N and longitude 8° 15' - 8° 40'E. It has a population of about 500,797 (The World Gazetteer, 2007), it lies in the Guinea savannah vegetative belt and on the bank of the river Benue which is the second largest river in Nigeria. The river divides the town into North and South banks and the town covers an area of 16 km² (Omudu and Amuta, 2007). It has a tropical climate with a temperature ranging between 21.70 -24.70⁰C (minimum) and 29.70-33.70⁰C (maximum). The climate of Makurdi town is the tropical wet and dry type, Koppen's Aw classification, with double maxima (Ayoade, 1983). The rainy season lasts from April to October, with five months of dry season (November to March). Annual rainfall in Makurdi town is consistently high, with an average annual total of approximately 1173 mm (Abah, 2012).

Data collection

Data of 1571 treated cases of CPVE in dogs presented at the VTH Annex, UAM from 2010 to 2016 were assessed. History, clinical signs and therapeutic regimens of CPV were reviewed. The presented signs were foul smelly haemorrhagic diarrhoea, emaciation, vomiting and lethargy. The vaccination history, treatment outcome, sex, breeds and age of dogs were recorded. The status of the vaccination history was categorized as vaccinated, unvaccinated and unknown. Similarly, the treatment outcomes of the cases following medical intervention were classified as either "recovered or dead". The suspected cases were confirmed using SensPERT[®] canine parvovirus antigen test kits (VetALL, Korea).

Statistical analysis

The data collected were analysed using Microsoft Office Excel (2007) for descriptive analysis of variables such as age, sex, and vaccination history and treatment outcome.

Ethical approval

This study was certified and approved and was performed according to the ethics of committee of the college of veterinary medicine, University of Agriculture, Makurdi, Nigeria.

RESULTS

Of the 1571 dogs that were brought to the VTH Annex, UAM, Makurdi, for over a period of seven years (2010 - 2016), 89 (5.7%) were diagnosed of CPV enteritis. Table 1 shows the analysis of diagnosed CPVE cases according to age, sex, vaccination status, treatment outcome and season where as the analysis of diagnosed CPVE cases by breed of dogs, month and year have been presented in diagrams 1-3. The local Nigerian breed had the highest frequency of cases 40 (44.9%), followed by Alsatian 18 (20.2%), Caucasian 7 (7.9%), Rottweiler, Russian shepherd 6 (6.7%), Mixed 4 (4.5%), Bull mastiff, Neapolitan mastiff 3 (3.4) and Pit-bull and Chihuahua 1(1.1%) (Diagram 1). The highest incidence of cases was recorded more in the month of January 22 (22.5%), June 14 (15.7%), March and July 11 (12.4%), February 8 (9%), May 7 (7.9%), April, October 4 (4.5%), December 3 (3.4%), September 2 (2.3%) and November 1 (1.1%) respectively (diagram 2) while the peak incidence of cases were recorded in the year 2012, 23 (25.8%), 2013, 16 (18%), 2014, 15 (16.9%), 2011, 11 (12.4%), 2015, 9 (10.1%), 2016, 8 (9%) and 2010, 7 (7.9%) respectively (Diagram 3).

Table 1. Analysis of diagnosed canine parvoviral enteritis cases base on age, sex, vaccination status treatment outcome and season, presented to the veterinary teaching hospital, university of agriculture, Makurdi from 2010 to 2016

Groups	Number of diagnosed cases of canine parvovirus enteritis	
	Frequency	Percentage (%)
Age (Months)		
0-5	71	79.8
6-12	12	13.5
>1 year	6	6.7
Sex		
Male	54	60.7
Female	35	39.3
Vaccination status		
Vaccinated	25	28.1
Unvaccinated	42	47.2
Unknown	22	24.7
Treatment outcome		
Recovered	76	85.4
Dead	13	14.6
Season		
Dry (Nov- March)	43	48.3
Wet (April- October)	46	51.7

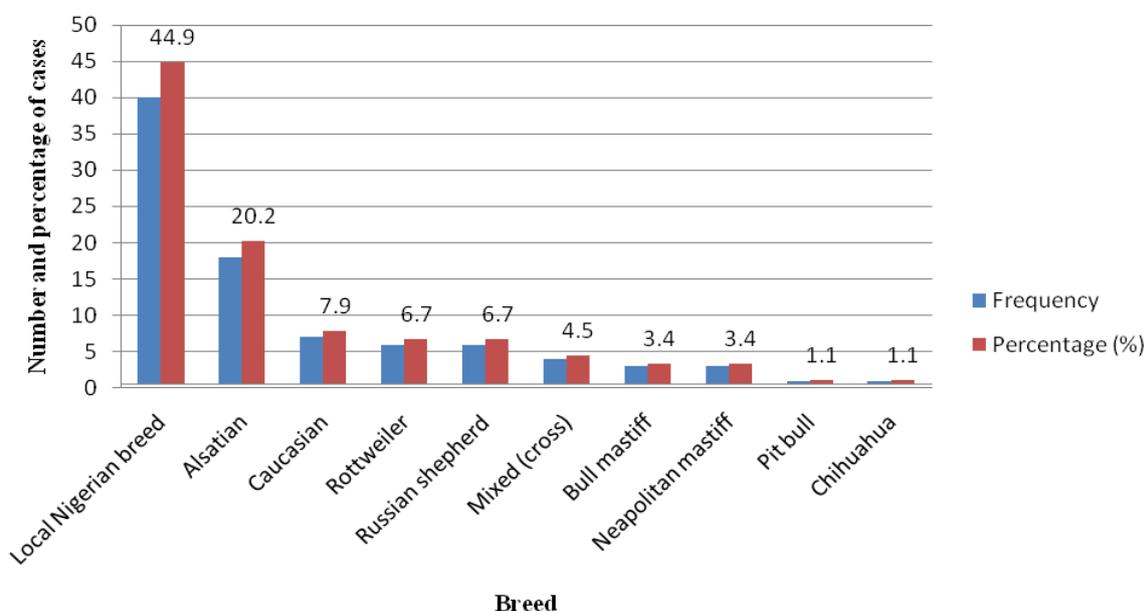


Diagram 1. The distribution of diagnosed canine parvoviral enteritis cases base on breed in university of agriculture Makurdi, Nigeria during 2010-2016.

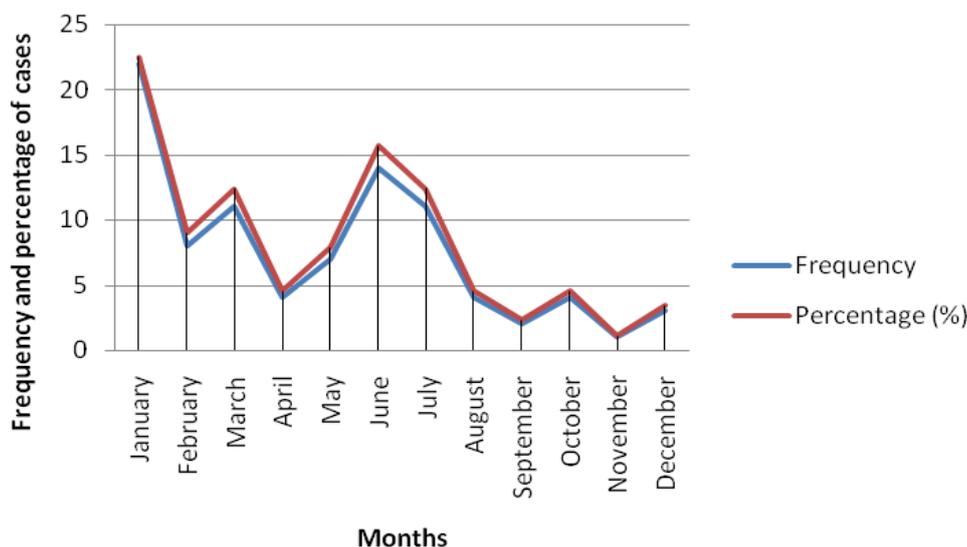


Diagram 2. The distribution of diagnosed canine parvoviral enteritis cases base on months in university of agriculture Makurdi, Nigeria during 2010-2016.

Frequency and percentage of cases of canine parvovirus enteritis

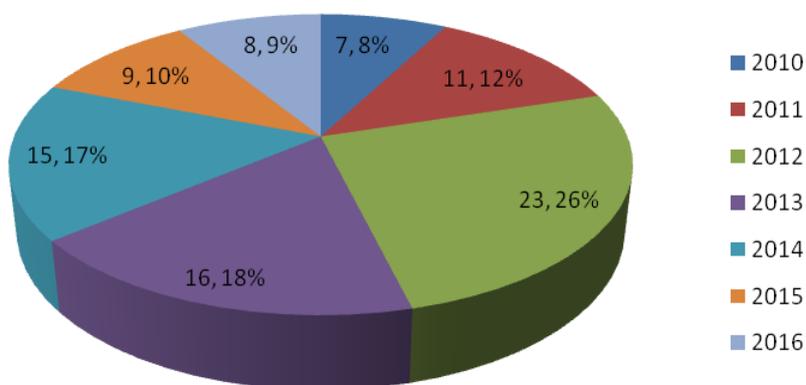


Diagram 3. The distribution of diagnosed canine parvoviral enteritis cases in Nigeria by years at the veterinary teaching hospital (2010-2016).

DISCUSSION

The study has shown that CPVE has been endemic in Makurdi metropolis for years with a 5.7 % prevalence rate (89/1571). Although, the prevalence rate in this study appears to be much lower compared to the values reported in Ilorin 6.4% (105/1645) (Daodu and Ajiboye, 2018), Effurun/Warri 13.4% (204/1527) (Shima et al., 2015), Jos North/South 17.4% (12/70) (Ogbu et al., 2016), Tunisia 32.1 % (54/168) (Tagorti, 2018), Nigeria and south Africa 96.7% (30/31), 98.2% (106/108) (Dogonyaro et al., 2013) and Zimbabwe 84.9% (191/225) (Mcree et al., 2014) but it was higher than the report from Vom 2.8% (87/3075) (Mohammed et al., 2005). The low prevalence may be due to low patronage of the VTH by dog owners, poor record keeping of medical cases and/or lack of proper diagnostic techniques/equipment then. Therefore, this report is a well representation of the cases of CPVE in Makurdi since the VTH is the major veterinary hospital where cases are handled in Makurdi metropolis.

The fact that the group (0-5 months) had a higher incidence rate of 79.8% (71/89), 6-12 months, 13.5% (12/89) and > 12 months, 6.7% (6/89) agrees with the reports from Effurun/Warri 60.3% (0-5 month), 27% (6-11 months) and 12.7% (12 months and above) (Shima et al., 2015), Argentina 86% (1-5 months) and 14% (6 months and above) (Calderon et al., 2011), United States of America 59% (less than 6 months) and 41% (more than 6 months) (Glickman et al., 1985), Slovenia 67.6% (1-5 months), 25.7% (6-12 months) and 6.8% (more than 12 months) (Gombac et al., 2008), India with 40.75% for (0-6 months), 24.19% for (7-12 months) and 10% for (12 months and above) (Basava, 2012). It has already

been reported that CPVE primarily affects puppies that are between the age of 6 weeks to 6 months (Marcovich et al., 2012; Mccaw and Hoskins, 2006; Prittie, 2004).

Local Nigerian breeds 44.9%, Alsations 20.2%, Caucasians 7.9%, Russian shepherd and Rottweiler 6.7%, mixed breed 4.5%, Neapolitan mastiff and Bull mastiff 3.4% each and the least Pit-bull and Chihuahua 1.1% each. The reasons for breeds of dog being susceptible to this dreaded disease remain unknown. Although, there have been earlier reports that Doberman Pinscher, Rottweiler and German Shepherd puppies are under greater risk of developing CPV enteritis than other breeds (Glickman et al., 1985; Houston et al., 1996; Castro et al., 2007; Shima et al., 2015). Our findings disagree with the report that Alsations, Rottweilers and Doberman pinschers appears to be more at risk (Gombac et al., 2008; Castro et al., 2007). Basava (2012) reported that, the Spitz breed had the highest prevalence (43.75%), followed by Doberman (42.85%), Pomeranian (37.5%), Alsatian (31.03%), Mongrel (28.71%) and Pug, Mastiff, Golden retriever, Labrador (27.77%). Dogs with highest incidence of cases of CPVE were the males (60.7%) as against females with (39.3%). This result agrees with the reported values in males (83.33%) and females (16.7%) respectively (Castro et al., 2007; Basava, 2012) but disagrees with the finding of Umar et al. (2015) who reported that female (58.5%) were more at risk than the males (41.5%). In Nigeria, male dogs are preferred to females by owners as security with exception of the few that breed them.

The prognosis of CPVE is as low as 9.1% in the absence of treatment, and 64% or higher with treatment (Otto et al., 1997). In the present study, the prognosis after treatment is 85.4%. This finding corroborates with the reports of Prittie (2004) and Macintire and Smith-Carr (1997) indicating that CPVE could be treated symptomatically. But unvaccinated dogs (47.2%) had the highest prevalence, followed by vaccinated dogs (28.1%) and dogs with unknown vaccination status (24.7%). This also agrees with the result of Basava (2012) who reported (35.4%) for unvaccinated and vaccinated dogs (16%) respectively. The lower prevalence rate of CPVE in vaccinated dogs indicated that current vaccine offer protection (Cavalli et al., 2001).

Monthly prevalence of CPVE in January (22.5%), June (15.7%), March and July (12.4%), February (9%), May (7.9%), April and October (4.5%), December (3.4%), September (2.3%) and November (1.1%) show that cases are seen in the dry season (48.36) and more in the wet season (51.7%), respectively. Shima et al. (2015) reported highest prevalence in January (17.2%) and lowest in April (2.0%). In contrast, Basava (2012) reported highest prevalence in July (48.97%) and the lowest in March (20%) and Houston et al. (1996) also reported that dogs are most likely to be admitted between July to September in Canada. Meanwhile, in the last seven years, the disease had its highest prevalence in 2012 (25.8%) followed by 2013 (18%), 2014 (16.1%), 2011 (12.4%), 2015 (10.1%), 2016 (8%) and 2010 (7.9%) respectively. This was due to lack of awareness on the preventive measures of the disease and poor regimen management.

CONCLUSION

CPVE is endemic in Makurdi metropolis and could be affected by age, sex, breed, vaccination status, treatment (management) and seasonal variations. Vaccination and therapy of the affected dogs could improve the well-being and longevity of the dogs. This result will serve as recorded information to veterinarians, dog owners and breeders in and around Makurdi in giving adequate care and taking preventive measures generally but particularly during the high prevalence period of CPVE.

DECLARATIONS

Author's contribution

ATM, AAA, NSA, IAF, AIW collated the data. NHC and TT analysed the data. TMT, ATT and SAS designed the work while TMT and SAS reviewed the manuscript.

Consent to publish

The author(s) grant(s) the publisher the sole and exclusive license of the full copyright in the contribution. Consequently, the publisher shall have the exclusive right throughout the world to publish and sell the contribution in all languages, in whole or in part, including, without limitation, any abridgement and substantial part thereof, in book form and in any other form including, without limitation, mechanical, digital, electronic and visual reproduction, electronic storage and retrieval systems, including internet and intranet delivery and all other forms of electronic publication now known or hereinafter invented.

Competing interests

The authors have declared that no competing interest exists.

REFERENCES

- Apa TT, Daly JM, and Tarlinton RE (2016). Canine parvovirus (CPV-2) variants circulating in Nigerian dogs. *Veterinary Record Open* 2016;3: e000198. Doi:10.1136/ vetreco-2016-000198.
- Ayoade JO (1983). Introduction to climatology for the tropics. Ibadan: Spectrum Books. pp. 179-184. Doi:<http://dx.doi.org/10.1144/GSL.JGS.1907.063.01-04.19>
- Bagshaw C, Isdell AE, Thiruvaiyaru DS, Brisbin JR IL and Sanchez S (2014). Molecular detection of canine parvovirus in flies (Diptera) at open and closed canine facilities in the eastern United States. *Preventive Veterinary Medicine*, 114: 276-284. Doi:<https://doi.org/10.1016/j.prevetmed.2014.02.005>.
- Basava RK (2012). Clinico-epidemiological studies on canine parvoviral infection in and around Tirupati. M. Sc. Thesis, Department of Veterinary Epidemiology and Preventive Medicine, Sri Venkateswara Veterinary University, Tirupati, India, pp 1-71.
- Bingga G, Liu Z, Zhang J, Zhu Y, Lin L, Ding S and Guo P (2014). High resolution melting curve analysis as a new tool for rapid identification of canine parvovirus type 2 strains. *Molecular and cellular probes*. Doi:<https://doi.org/10.1016/j.mcp.2014.08.001>.
- Bourn D, Wint W, Blench R and Woolley E (1994). Nigerian Livestock Resources Survey. *World Animal Review*, 78: 49-58.
- Brown AJ and Otto CM (2008). Fluid therapy in vomiting and diarrhea. *Veterinary Clinics of North America: Small Animal Practice*. 38(3):653-75. Doi: 10.1016/j.cvsm.2008.01.008.
- Buonavoglia C, Martella V, Pratelli A, Tempesta M, Cavalli A, Buonavoglia D, Bozzo G, Elia G, Decaro N and Carmichael L (2001). Evidence for evolution of canine parvovirus type 2 in Italy. *Journal of General Virology*, 82: 3021-3025. Doi: 10.1099/0022-1317-82-12-3021.
- Calderón MG, Romanuttia C, D'Antuonoa A, Kellerb L, Mattiona N and La Torre J (2011). Evolution of canine parvovirus in Argentina between years 2003 and 2010: CPV2c has become the predominant variant affecting the domestic dog population. *Virus Research*. 157:106-110. Doi:<https://doi.org/10.1016/j.virusres.2011.02.015>.
- Carmichael LE (1994). Canine parvovirus type-2: An evolving pathogen of dogs. *Annals of Veterinary Medicine*. 135(4): 590-464.
- Carmichael LE (2003). Canine infectious Diseases - A personal perspective. An oral presentation given on August 16th 2003, at the International Symposium: "Reunion Mundial de Lideres en la Educacion Veterinaria" that commemorate the 150th anniversary of veterinary education in the College of Veterinary Medicine, National Autonomous University of Mexico, Mexico City, D.F. Baker Institute for Animal Health Cornell University, Ithaca, New York (USA).
- Castro TX, Miranda SC, Labarthe NV, Silva LE and Cubel Garcia RCN (2007). Clinical and Epidemiological Aspects of Canine Parvovirus (CPV) Enteritis in the State of Rio de Janeiro: 1995-2004. *Arquivo Brasileiro de Medicina Veterinariae Zootecnia*, 59: 333-339. Doi:<http://dx.doi.org/10.1590/S0102-09352007000200010>.
- Cavalli A, Bozzo G, Decaro N, Tinelli A, Aliberti A and Buonavoglia D (2001). Characterization of a canine parvovirus strain isolated from an adult dog. *New Microbiology*, 24: 239-242.
- Chollom S, Fyaktu E, Okwori A, Agada G, Hashimu G, Akele R, Voumangai E, Dashe T and Egah D (2013). Molecular detection of canine parvovirus in Jos, Nigeria. *Journal of Veterinary Medicine and Animal Health*, 5: 57-59. DOI:10.5897/JVMAH12.033.
- Cynthia MK and Scott L (2010). *The Mercks Veterinary Manual*, Tenth edition. Merck and Co Inc, White House Station, N. J. USA.
- Daodu, O.B., Amosun, E.A. and Oluwayelu, D.O. (2017). Antibiotic resistance profiling and microbiota of the upper respiratory tract of apparently healthy dogs in Ibadan, South west Nigeria. *African Journal of Infectious Disease*, 11 (1): 1-11 Doi:10.21010/ajid.v11n1.
- Decaro N, Elia G, Martella V, Desario C, Campolo M, Trani LD, Tarsitano E, Tempesta M and Buonavoglia C (2005b). A real-time PCR assay for rapid detection and quantitation of canine parvovirus type 2 in the faeces of dogs. *Veterinary microbiology*, 105: 19-28. DOI: 10.1016/j.vetmic.2004.09.018.
- Desario C, Decaro N, Campolo M, Cavalli A, Cirone F, Elia G, Martella V, Lorusso E, Camero M and Buonavoglia C (2005). Canine parvovirus infection: which diagnostic test for virus? *J. Virol. Methods*, 126(1): 179-185. DOI: 10.1016/j.jviromet.2005.02.006.
- Dogonyaro BB, Bosman AM, Sibeko KP, Venter EH and Van Vuuren M (2013). Genetic analysis of the VP2-encoding gene of canine parvovirus strains from Africa. *Veterinary microbiology*, 165: 460-465. Doi:<https://doi.org/10.1016/j.vetmic.2013.04.022>
- Eghafona N, Jacob J and Yah S (2007). Evaluation of post-vaccination immunity to canine distemper and parvoviruses in Benin City, Nigeria. *African Journal of Biotechnology* 6: 1898-1904. Doi: <http://dx.doi.org/10.5897/AJB2007.000-2286>
- Ezeibe MC, Nwaogu IC, Nwigwe AN, Okorafor ON and Eze JI (2010). Aluminium-magnesium silicate inhibits parvovirus and cures infected dogs. *Health*, 2: 1215. Doi:10.4236/health.2010.210179
- Ezeokoli CD, Umoh JU, Adeyanju JB and Abdullahi SU (1985). Parvovirus enteritis in Nigerian dogs. *Journal of Small Animal Practice*, 26: 669 - 673. Doi:<https://doi.org/10.1111/j.1748-5827.1985.tb02194.x>.
- Glickman LT, Domanski LM, Patronek GJ and Visintainer F (1985). Breed-related risk factors for canine parvovirus enteritis. *Journal of American Veterinary Medical Association*, 187(6): 589-594.
- Gombac M, Svava T, Tadic M and Pogacnik M (2008). Retrospective Study of Canine Parvovirus in Slovenia: Case Report. *Slovenia Veterinary Research*, 45: 73-78.
- Hoskins JD (1997). Update on canine parvoviral enteritis. *Veterinary Medicine*, 92(8):694-709.
- Houston DM, Ribble CS and Head LL (1996). Risk factors associated with parvovirus enteritis in dogs: 283 cases (1982-1991). *Journal of the American Veterinary Medical Association*, 208(4):542-546.

- Oh JS, Ha GW, Cho YS, Kim MJ, An DJ, Hwang KK, Lim YK, Park BK, Kang B and Song DS (2006). One-step immunochromatography assay kit for detecting antibodies to canine parvovirus. *Clinical and Vaccine Immunology*, 13(4):520–4. Doi: 10.1128/CVI.13.4.520-524.2006.
- Lamm CG and Rezabek GB (2008). Parvovirus infection in domestic companion animals. *Veterinary Clinics of North America: Small Animal Practice*, 38(4): 837–50. Doi: 10.1016/j.cvsm.2008.03.008.
- Macintire DK and Smith-Carr S (1997). Canine parvovirus. Part II. Clinical signs, diagnosis, and treatment. *Compendium on Continuing Education for the Practising Veterinarian*, 19(3):291–302.
- MacLachlan NJ and Dubovi EJ (2011). Rabies. In: Fenner's *Veterinary Virology*. 4th edition, pp 327-336.
- Marcovich JE, Stucker KM, Carr AH, Harbison CE, Scarlett JM and Parrish CR (2012). Effects of canine parvovirus strain variations on diagnostic test results and clinical management of enteritis in dogs. *Journal of the American Veterinary Medical Association*, 241(1):66–72. Doi:10.2460/javma.241.1.66.
- Martella V, Cavalli A, Decaro N, Elia G, Desario C, Campolo M, Bozzo G, Tarsitano E and Buonavoglia C (2005). Immunogenicity of an intranasally administered modified live canine parvovirus type 2b vaccine in pups with maternally derived antibodies. *Clinical and Diagnostic Laboratory Immunology*, 12(10): 1243–1245. Doi:10.1128/CDLI.12.10.1243-1245.2005.
- McCaw DL, Hoskins JD (2006). Canine viral enteritis. In: Green CE, editor. *Infectious Diseases of the Dog and Cat*. 4th ed. St Louis, MO: Saunders; pp 63–73.
- Mcree A, Wilkes RP, Dawson J, Parry R, Foggin C, Adams, H., Odoi A and Kennedy AM (2014). Serological Detection of Infection with Canine Distemper Virus, Canine Parvovirus and Canine Adenovirus in Communal Dogs from Zimbabwe. *Journal of the South African Veterinary Association*, 85(1), Art. #1110, 2 p. Doi:http://dx.doi.org/10.4102/jsava.v85i1.1110.
- Mohammed JG, Ogbe AO, Zwandor NJ and Umoh JU (2005). Risk factors associated with canine parvovirus enteritis in Vom and environs. *Animal Research International*. 2(3): 366 – 368. http://dx.doi.org/10.4314/ari.v2i3.40870
- Muzyczka N, Berns KI (2001). Parvoviridae: The Viruses and Their Replication. In: Knipe DM, Howley PM, Griffen DE, Lamb RA, Martin MA, Roizman B and Straus ES (Eds.), *Fields Virology*. 4th ed. Lippincott Williams & Wilkins, Philadelphia, PA, pp. 2327–2359.
- Mylonakis ME, Kalli I and Rallis TS (2016). Canine parvoviral enteritis: an update on the clinical diagnosis, treatment, and prevention. *Veterinary Medicine: Research and Reports*, 7: 91—100. Doi: https://doi.org/10.2147/VMRR.S80971.
- Nivy R, Hahn S, Perl S, Karnieli A, Karnieli O and Aroch I (2011). A Fatal Outbreak of Parvovirus Infection: First Detection of Canine Parvovirus Type 2c in Israel with Secondary Escherichia coli Septicemia and Meningoencephalitis. This year we celebrate 250 years of veterinary medicine. The world's first veterinary school was established in Lyon, France in 1761 by Claude Bourgelat who managed to persuade King Louis XV of France of the need to train specialists, 66: 3.
- Nwoha RIO (2011). Parvoviral Enteritis in a Dog: Case Report and Review of the Literature. *Continental Journal of Veterinary Science*, 5: 6-10.
- Ogbu KI, Chukwudi IC, Ijomanta OJ, Agwu OE and Chinonye CN (2016). Prevalence of Canine Parvovirus in Jos North and South Local Government Areas of Plateau State. *British Microbiology Research Journal*, 13(2): 1-5. DOI: 10.9734/BMRJ/2016/22813.
- Omudu EA and Amuta EU (2007). Parasitology and urban livestock farming in Nigeria: Prevalence of ova in faecal and soil samples and animals ectoparasites in Makurdi. *Journal of the South African Veterinary Association*, 78: 40-45.
- Otto CM, Drobatz KJ, Soter C (1997). Endotoxemia and tumor necrosis factor activity in dogs with naturally occurring parvoviral enteritis. *Journal of Veterinary Internal Medicine*, 11(2): 65–70.
- Parker JSL, Murphy WJ, Wang D, O'Brein SJ and Parrish CR (2001). Canine and feline parvoviruses can use human and feline transferrin receptors to bind, enter and infect cells. *Journal of Virology*, 75: 3896-902. DOI: 10.1128/JVI.75.8.3896-3902.2001.
- Pollock RVH and Carmichael LE (1982). Maternally Derived Immunity to Canine Parvovirus Infection: Transfer, Decline and Interference with Vaccination. *Journal of the American Veterinary Medical Association*, 180(1): 37-42.
- Pollock RVH and Carmichael LE (1988). Canine viral enteritis. In: Barlough JE, editor. *Manual of small animal infectious diseases*. New York: Churchill Livingstone, pp 101–7.
- Prittie J (2004). Canine parvoviral enteritis: a review of diagnosis, management, and prevention. *Journal of Veterinary Emergency and Critical Care*, 14(3):167-176. Doi: https://doi.org/10.1111/j.1534-6935.2004.04020.x
- Shima F, Aapa, T and Mosugu JT (2015). Epidemiology of Canine Parvovirus Enteritis among Hospitalized Dogs in Effurun/Warri Metropolitan Region of Delta State, Nigeria. *Open Access Library Journal*, 2: 1-7. Doi:http://dx.doi.org/10.4236/oalib.1101208.
- Smith-Carr S, Macintire DK, Swango LJ (1997). Canine parvovirus. Part I. Pathogenesis and vaccination. *Compendium on Continuing Education for the Practising Veterinarian*. 19(2):125–133.
- Tagorti G (2018). Prevalence of canine parvovirus infection in Grand Tunis, Tunisia. *Journal of Advanced Veterinary and Animal Research*. 5(1):93-97. Doi: http://doi.org/10.5455/javar.2018.e251.
- The World Gazetteer (2007). Accessed via: http://mikes.railhistory.railfan.net.
- Touhri L, Bouzid I, Daoud R, Desario C, El Goulli AF, Decaro N, Ghorbel A, Buonavoglia C and Bahloul C (2009). Molecular characterization of canine parvovirus-2 variants circulating in Tunisia. *Virus genes*, 38: 249-258. https://doi.org/10.1007/s11262-008-0314-1.
- Umar S, Ali A, Younus M, Maan MK, Ali S, Khan WA and Irfan M. (2015). Prevalence of Canine Parvovirus Infection at Different Pet Clinics in Lahore, Pakistan. *Pakistan journal of zoology*. 47(3):657-663.
- Wilson S, Illambas J, Siedek E, Stirling C, Thomas A, Plevova E, Sture G and Salt J (2014). Vaccination of dogs with canine parvovirus type 2b (CPV- 2b) induces neutralising antibody responses to CPV-2a and CPV-2c. *Vaccine*, 32: 5420- 5424. DOI: 10.1016/j.vaccine.2014.07.102.



Application of Different Hormonal Protocols for Improving Reproductive Performance of Barki Ewes

Gamal Ashour¹, Moharram Fouad El-Bassiony^{2*}, Sherif Mohamed Dessouki¹ and Mohamed Awad El-Wakeel²

¹Department of Animal Production, Faculty of Agriculture, Cairo University, 12613, Giza, Egypt

²Animal and Poultry Physiology Department, Desert Research Center, 11753, Cairo, Egypt

*Corresponding author's Email: moharramf@yahoo.com

ABSTRACT

This investigation was designed to assess reproductive improvement in Barki ewes using different hormonal protocols. Seventy-five non-pregnant and non-lactating Barki ewes were randomly assigned into three equal groups (25 ewes each), namely G1, G2 and G3. (G1) served as control, while (G2) was treated with an intravaginal progestagen impregnated sponge for 12 days then was removed. On the ninth day, all treated ewes received an intramuscular injection of prostaglandin F_{2α}. (G3) was also synchronized as G2, in addition to on the 10th day ewes were received an intramuscular injection of 750 IU PMSG in descending doses for three days so that the last dose was injected at the 12th day concurrently with sponges' removal. Meanwhile, on the 14th day, all ewes in G3 were received an intramuscular injection of hCG hormone (500 IU/ewe). The results showed an increase in plasma progesterone level from the first day of pregnancy and rises up to the last day before parturition. Progesterone was found to be higher in G3 (that had higher twinning rate) than in G2 than in G1. Insignificant increase in estrus response was observed in groups (G2, G3) compared to G1 (100, 100 and 92%, respectively). Conception rate was significantly higher in G3 (100%) than G2 (92%) and G1 (88%). While, abortion rate was higher in G1 and surpassing G2 and G3 (9.0, 0.0 and 0.0 %, respectively). Lambing rate was significantly higher in G3 as compared to G2 and G1 groups (100, 92 and 80 %, respectively). While, weaning rate was recorded to be insignificantly higher in G2 than in G1 and G3 (100.0, 95.45 and 94.29%, respectively). It could be concluded that, hormonal manipulation using intravaginal progestagen impregnated sponge and PMSG in the presence of hCG; would be a proper way for enhancing the reproductive efficiency of Barki ewes.

Key words: Barki ewes, Synchronization, Superovulation, Twinning, Reproduction

INTRODUCTION

Low reproductive performance has been observed in sheep flocks raised traditionally in arid regions. This poor performance may be attributed to several factors including fertility, selection, nutrition and disease (Webb et al., 2004; Yavuzer, 2005). Increasing Barki sheep productivity by increasing lambing frequency and fecundity is considered to be the most important factor in the development of Barki sheep production. In this regard, increasing the rate of fecundity in sheep offers the best opportunity to increase the efficiency of lamb meat production (Koyuncu and Alticekic, 2010). Natural lambing in Barki sheep occurs throughout the year in a scattered manner negatively affecting survival and growth rates of the lambs born during the unfavorable season of the year. Consequently, one of the most common ways to alleviate problems related to disorderly lambing is the using of exogenous source of hormones for artificially controlling the time of mating (Mekuriaw et al., 2016). In addition, the use of hormonal manipulation for estrus synchronization creates the opportunity for timed breeding and lambing. This in turn results in taking the advantages of seasonal variation of forage availability, photoperiod, labor resources and market demands. Estrus synchronization is a valuable management tool that has been employed successfully in enhancing reproductive efficiency, particularly in cows, ewes and does as early reported by Kusina et al. (2000). The success of a hormonal treatment for estrus synchronization depends on the productive status of ewes, season of the year, the type of hormonal protocol and the method of administration. Multiple ovulation programs are also possible with the use of estrus synchronization for improving herd productivity.

Estrus synchronization in small ruminants is achieved either by reducing the length of the luteal phase of the estrous cycle with prostaglandin F_{2α} or by artificially extending it with exogenous progesterone or potent progestagens (Jainudeen et al., 2000 and Kusina et al., 2000). Synchronization of estrus has been practiced for the last few decades when many protocols varying in degrees of success which based on the use of intravaginal progestagen devices inserted for 7 to 14 days followed by gonadotropin injection at the time of devices' removal and introduction of teaser ram

ORIGINAL ARTICLE
 pii: S232245681800006-8
 Received: 28 July 2018
 Accepted: 10 Aug 2018

(Zarkawi, 2001). Also, hormonal treatments to control ovulation and reproduction are a prerequisite for successful breeding and increasing the number of pregnant females (Motlomelo et al., 2002 and Husein et al., 2005). In small ruminants, several programs have been used for improving fertility by controlling the ovarian activity. Acceptable results of estrus responses have been achieved by using progestagen and equine Chorionic Gonadotropin (eCG) together at the time of progestagen withdrawal (Fonseca et al., 2005; Husein et al., 2005).

Application of assisted reproductive technologies is a prerequisite for improving reproductivity and productivity of livestock, especially in arid and semi-arid environments species which are usually characterized by low fertility. Most recently, Kaya et al. (2018) noted that hormonal protocols are one of the main items to improve, herds productivity including superovulation as proposed by Souza et al. (2014). In practice, generality of the superovulation protocols used for embryo production in ewes, both *in vivo* and *in vitro*, consists of the administration of intravaginal progestagen impregnated sponges; both for synchronization of estrus and ovulation or for avoidance of spontaneous ovulation (Cognie et al., 2003). The objectives of superovulation included inducing a high number of ovulations and subsequent high fertilization rate, while at the same time ensuring a normal physiological environment in the reproductive tract for embryo development. The application of superovulation technology to improve endogenous secretions of pregnant hormones increases offspring productivity and gross revenue in the small scale farm (Andriyanto and Manalu, 2011).

The present study was conducted to place on record the lambing rates of ewes following and to investigate the possibility of improving fertility and increasing twinning rate in Barki ewes under arid conditions of the North Western Coast of Egypt using different hormonal protocols for estrus synchronization and superovulation.

MATERIALS AND METHODS

Experimental region

The present study was carried out during the period from June 2015 to January 2016 at Animal Production Unit, Sustainable Development Center for Matrouh Resources, Matrouh Governorate, which belongs to the Desert Research Center (DRC), Ministry of Agriculture and Land Reclamation. The station is located at 240 Km West of Alexandria and 222 Km Egyptian western borders.

Ethical approval

This experiment was performed according to all ethics and animal rights (Desert Research Center). As much as this work had considering all rules and regulations in conformity with the European union directive for the protection of experimental animals (2010/63/EU).

Experimental animals, feeding and management

Seventy-five, non-pregnant and non-lactating, Barki ewes averaged 48.10 ± 0.72 kg Live Body Weight (LBW) and ranged between 2.5 and 3.5 years old were used in this study. All ewes were clinically examined for any reproductive disorders as well as general health status. Vaccination against the major prevailing epidemic diseases, internal and external parasites were controlled in proper time. Ewes were weighed before starting the experiment, and kept under an intensive production system and housed in semi-open yards throughout the experimental period. Lambs were left all the day time with their dams for suckling up to weaning age at three months of age. All groups were daily fed on a concentrate (0.75 kg) and berseem hay (0.5kg) per head during the experimental period to cover their nutrient requirements during different physiological status according to Kears (1982). Lambs were daily fed only on their dams' milk from birth to weaning age at three months of age. The daily ration was offered in a certain mode of feeding starting with concentrate mixture at 08:30 h followed by chopped rice straw at 12:00 h and continuing to the next morning feeding. All animals drank fresh water three times a day (08:00, 14:00 and 20:00 h).

Experimental design

Animals were randomly divided into three equal groups (25 ewes per group (Figure 1). The first group (G1) was served as a control without any hormonal treatment and was left without synchronization or superovulation. While, the second group (G2) was treated using an intravaginal progestagen impregnated sponge (20 mg cronolone, Chronogest® CR, product of Intervet international B. V. and manufactured in the European Union, (EU) was inserted. Sponges remained in situ for 12 days and were removed on the 12th day. On the ninth day, all ewes were received an intramuscular injection of prostaglandin $F_{2\alpha}$ (1 ml Synchronate each 1 ml solution containing 0.250 mg Cloprostenol, Bremer Pharma GMBH, Germany). The third group (G3) was also synchronized using an intravaginal progestagen impregnated sponge for 12 days and received an intramuscular injection of prostaglandin $F_{2\alpha}$, on the ninth day, while on the 10th day ewes of the third group (G3) were received an intramuscular injection of 750 IU PMSG (Laboratorios Hipra, S.A. Avda. Ia Selva, 135, 17170 Amer (Girona) Spain). PMSG injections were applied in gradual decreasing doses for

three days that each ewe received 275 IU at the 10th day, the second dose (250 IU) was injected at the 11th day, while the last dose (225 IU) was injected at the 12th day at the time of sponges' removal. Meanwhile, on the 14th day, all ewes in G3 group received an intramuscular injection of 500 IU/ewe of hCG hormone (Epifasi lyophilized ampoule contains 5000 I.U. of Human Chorionic Gonadotrophin, hCG) and 10 mg of lactose, manufactured by Egyptian int. pharmaceutical industries Co. (EIPICO, Egypt). During the synchronization period, all synchronized animals were subjected to a twice daily check (morning and evening) to ensure that sponges remained in their position during the treatment period.

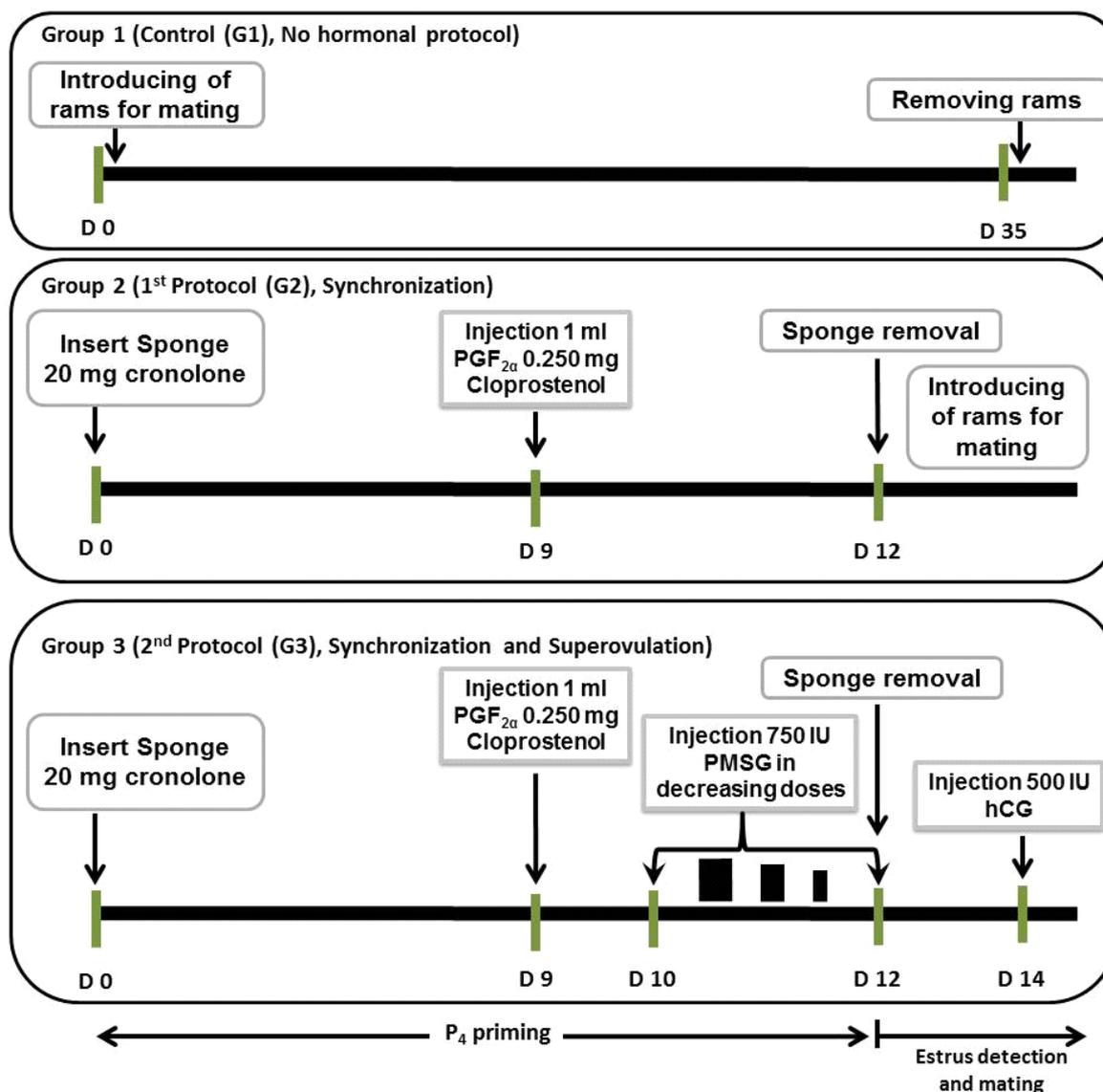


Figure 1. Experimental design and timeline for the treatment administration for the control (G1), synchronization (G2) and synchronization + superovulation (G3) groups.

Blood sampling and progesterone assay

Blood samples (5 ml) were collected from the jugular vein into EDTA (ethylenediamine tetra-acetic acid) containing tubes. Blood samples were collected immediately before sponge insertion on day 0 from ewes. The rest of the samples were withdrawn on days 1, 3, 5, 7, 9, 10, 12, 13, 15, 17, 19, 30, 60, 90, 120, 150, 180 from all groups. Blood plasma was pipetted into 1.5 mL Eppendorf tubes using sterilized plastic disposable Pasteur pipettes and then stored at -20 °C until assayed for (P₄) concentration. Progesterone hormone was quantified by ELISA method using BIOS kit provided by Chemux Bioscience Corporation, 385 Oyster Point Blvd Suite 5-6., South San Francisco, CA 94080, USA. The standard curve ranged between 0-50 ng/ml. The sensitivity of the curve was 0.2 ng/ml.

Estrus detection and mating

Nine fertile Barki rams were introduced to the ewes of all experimental groups, three rams for each group for estrus detection and mating; started 24 hours after removing the sponges (G2 and G3) and left to one week. While the control group (G1), rams left with ewes from the zero day throughout two estrus cycles (35 days total). Rams were allowed to

interchange among different ewe groups to avoid sire/group confounding effect. The fertile rams' chest was colored in a different color for each group.

Rams were introduced to ewes for seven days and left with them for estrus detection and natural mating. Estrus was checked continuously by observation of the color marks on the ewe's rumps. Ewes were considered in estrus and mating had occurred when the paint mark was heavy and evenly distributed or copulation was observed.

Reproductive parameters

The following traits were estimated for each of the treated groups:

- Estrus response = number of ewes showing signs of estrus / total ewes treated in each group ×100.
- Conception rate = number of ewes conceived / number of ewes showing estrus and mated in each group ×100.
- Lambing rate = number of ewes lambed / number of ewes mated in each group ×100.
- Weaning rate = number of lambs weaned / number of lambs born in each group ×100.
- Abortion rate = number of ewes aborted / number of ewes conceived in each group ×100.
- Litter size at birth = number of lambs born / number of ewe lambed.
- Litter size at weaning = number of lambs weaned / number of ewe lambed.
- Average litter weight at birth = the kilograms lambs born / number of ewe lambed.
- Fecundity rate = number of lambs born / number of ewe mated.
- Weaning weight = the kilograms lambs weaned / number of lambs weaned.
- Mortality rate up to weaning (%) =live lambs born - lambs weaned / live lambs born x100.

Statistical analysis

A General Linear Model procedure (SAS, 2004) was used for the statistical analyses of progesterone concentration, milk yield and composition during the experimental period using the following model:

$$Y_{ijk} = \mu + P_i + T_j + (P*T)_{ij} + e_{ijk}$$

Where,

- Y_{ijk} = Any observation of k^{th} animal within j^{th} treatment within i^{th} period
- μ = Overall mean
- P_i = Effect of i^{th} period
- T_j = Effect of j^{th} treatment ($j = 1-3$, 1= Tr1, 2= Tr2, 3= Tr3)
- $(P*T)_{ij}$ = The interaction between period and treatment
- e_{ijk} = Experimental error

Significant differences among means were detected using Duncan's multiple range test (Duncan, 1955). While, reproductive traits (estrus response, conception, lambing, weaning, fecundity, abortion, single, twinning, triple, mortality rates) and litter size at birth, litter size at weaning, weaning weight, sex ratio were analyzed using Chi-square test.

RESULTS AND DISCUSSION

Estrus response, and conception and abortion rates

The data presented in table (1) revealed that estrus response was non-significantly high in G2 (FGA+PGF2 α) and G3 (FGA+PGF2 α +PMSG and hCG) than the G1 (control group). The recorded data were (100, 100 and 92%, respectively). The data also indicated that conception rate was significantly higher ($P<0.05$) in G3 than in G2 and G1 (100, 92 and 88%). On the other hand, abortion rate was higher in control group (G1) surpassing G2 and G3, as both groups G2 and G3 showed no abortion (9.0, 0.0 and 0.0%, respectively) (Table 1).

Table 1. Estrus response and conception and abortion rates % of Barki ewes as affected by different hormonal protocols (LSM \pm SE)

Traits	Treatment (T)			Overall mean	\pm SEM	Significance
	G1	G2	G3			
Estrus response	92.00 (23/25)	100.0 (25/25)	100.0 (25/25)	97.33 (73/75)	5.30	NS
Conception rate	88.00 (22/25)	92.00 (23/25)	100.00 (25/25)	93.33 (70/75)	4.87	*
Abortion rate	09.00 (2/22)	00.00	00.00			

G1 = control group; G2 = FGA and PGF2 α ; G3 = FGA, PGF2 α , PMSG and hCG; NS = non-significant

The results of this study confirm that using the intravaginal progestagen impregnated sponge is an effective tool for the synchronization of the estrus. These results were previously confirmed in goats at the beginning of the breeding season in semi-desert conditions (Riaz et al., 2012). In our study, progesterone treatment lasted for 12 days showed high efficiency. Hosseinipناه et al. (2014) concluded that the occurrence of estrus using CIDR in non-reproductive season for 10, 12 and 14 days was associated with the onset of good estrus and use of GnRH had positive effect on fertility in treatment groups, with the highest significant value for the 12 days P4 treatment. In the same year, Zoharaa et al. (2014) recorded higher pregnancy rates as observed in the 30 mg FGA treatment (100%) compared with 100 µg Cloprostenol (88.2 %), 175 µg Cloprostenol (75%) and 45 mg FGA (93.8 %) treatments. It was concluded that though FGA sponge protocol presented superior results.

In comparison with our results, Abdul Muin et al. (2013) suggested that CIDR treatment for 14 days with 400 IU PMSG and 0.05 mg cloprostenol prior to CIDR removal gave better result in estrus synchronisation than in CIDR treatment for 9 days with 0.05 mg cloprostenol given 24 hours before CIDR removal. In the present study both treatments were high to estrous response with no significant except to the control group. But the results of this study agree with recent report of Waheeb et al. (2017) who concluded that intravaginal progesterone sponge for 7 days+500IU eCG at sponge removal is convenient for estrus synchronization of ewes raised in field conditions during breeding season specially that the reproductive and fertility parameters recorded in his study were acceptable and within values reported previously in farm conditions. Also, Dendena (2017) stated a fertility rate of 85.1% after the administration of 500 IU of eCG, which was considered lower than PMSG treatment in this study, this may be referred to using higher dose (750 IU) in our treated group. Thus, our results were comparable to what has been most recently reported by Fornazari et al. (2018) as these stated that short-term progestagens (FGA e MAP) + eCG were 100.0% efficient in Assaf breed in cycle control worth to mention that in the same study the administration of 750 IU of eCG determined a fertility rate of 76.5%, which was controversially to our results. From our point of view, less prolific and less seasonal breeds tend to be more responsive to eCG administration, specially that different follicular populations present on the ovaries before progestagen treatments + eCG may also condition fertility rate (Omontese et al., 2016). In general, the present results meet these last investigators' observations that exogenous gonadotropin administration advanced ovulation and higher estrus synchronization precision (Letelier et al., 2011; Valentim et al., 2016), as they support ovarian mechanisms affecting follicular growth and maturation and promoting the proper luteinization of the CL (Valentim et al., 2016). In most studies, it is mentioned that in adult animals the expected estrus rate is greater than 90%, while nulliparous goats can reach up to 97.2% and lactating animals up to 85.7% (Abecia et al., 2011; Navanukraw et al., 2014; Alvarado-Espino et al., 2016).

Lambing rate, abortion rate, weaning rate, fecundity rate, litter size at birth, litter size at weaning and mortality rates (%)

Lambing rate is expressed as the percentage of ewes lambing out of ewes mated. As shown in **table 2** lambing rate was significantly higher ($P < 0.05$) in G3 as compared to G2 or G1 (100, 92 and 80%, respectively). While, weaning rate was recorded to be higher in G2 than in G1 and G3 and showed non-significantly rates of 100.0, 95.45 and 94.29%, respectively.

Table 2. Lambing rate, weaning rate, fecundity rate, litter size at birth, weaning rate, litter size at weaning and weaning weight of Barki ewes as affected by different hormonal treatment (LSM±SE)

Traits	Treatment (T)			Overall mean	± SEM	Sig.
	G1	G2	G3			
Lambing rate (%)	80.00 (20/25)	92.00 (23/25)	100.00 (25/25)	90.67 (68/75)	4.63	**
Weaning rate (%)	95.45 (21/22)	100.0 (26/26)	94.29 (33/35)	96.39	5.95	NS
Fecundity rate (%)	0.88 ^b (22/25)	1.04 ^b (26/25)	1.40 ^a (35/25)	1.10	0.10	**
Mortality rate (%)	4.55 (1/22)	0.00 (0/26)	5.71 (2/35)	3.61	--	NS
Litter size at birth/ewe	1.10 ^b (22/20)	1.13 ^b (26/23)	1.40 ^a (35/25)	1.22	0.05	*
Litter size at weaning/ewe	1.05 (21/20)	1.13 (26/23)	1.32 (33/25)	1.17	0.05	NS
Day 90 (Weaning weight)(kg)	16.71	17.53	17.45	17.28	--	NS

G1 = control group; G2 = FGA and PGF2α; G3 = FGA, PGF2α, PMSG and hCG; Sig. = significance; NS = non-significant

Means within each row with different superscripts are significantly different at 5% level.

As reported in our study, fecundity rate was significant higher in G3 and G2 than in G1 (1.40, 1.04 and 0.88), respectively (**Table 2**). Moreover, litter size at birth as expressed as number of born lambs per ewe lambled is considered to be an important parameter of fertility. Data illustrated in table 2 indicated that litter size at birth was significantly the highest in G3 followed by G2 as compared to G1 (1.40, 1.13 and 1.10, respectively). Litter size at weaning was significantly higher in G3 then G2 than in G1 (1.32, 1.13 and 1.05 respectively). The data also revealed that weaning weight showed insignificant increase in G2 as compared to G3 and G1 (17.53, 17.45 and 16.71, respectively). Non-significant higher values were recorded for mortality rate in G3 and G1 than in G2 (5.71, 4.55 and 0.00%, respectively).

In the tropics, biological and economic efficiency of animal production are predominantly determined by reproductive performance. Fertility is categorized as important parameters for determination the productivity of sheep; fertility may be estimated by several indicators as litter size per lambing for example which is more important than weight gain of lambs. Profit gain from increasing lamb survival is the major concern of sheep producers (Elliott et al., 2011). According to Elliott et al. (2011), sheep producers believed that using teasers for estrus synchronization, minimizing handling and interrupting ewes during the peri-partum period, and limiting duration of the breeding period would be beneficial to lamb survival. Contrarily to our results, it has been confirmed that high dose of eCG increases the ovulation rate but also reduces the embryonic survival rate (Diskin and Morris, 2008).

Formerly Gongnet (1996) reported malnutrition as the main cause of lamb mortality before weaning in lambs, for that reason, in recent study by Allou et al. (2013) advised with proper diet to enhance the adequate health that improve the survivability and lambs growth during the first three months of age. While, Gbangboche et al. (2005) referred the lamb survival rate before weaning to the age of the sheep at the first lambing influences, this was confirmed by Allou et al. (2013), who had stated that lambs survival is positively and beneficially correlated with this influence. Early reports indicated that the number of lambs surviving until weaning has been found to be influenced by ewes' body condition score at mating (Carson et al., 2001) rather than other surrounding factors.

Lambs with intermediate birth weights have a lower mortality risk than small and very large lambs (Piwczynski et al., 2012). However, the association between lamb birth weight and mortality was not detected in a study by Brien et al. (2009). In this regard, due to negative effects of large litters on lamb survival, close supervision should be provided to non-singleton lambs (Chniter et al., 2011). Litter size is influenced by several factors, and is low in ewes younger than 2 years of age and ewes having low weight at mating (Atashi et al., 2013). Weight gain in lambs is described by two subsequent phases, the first lasts from birth up to 90 days with a moderate weight change, this phase is followed by very fast changes in weight form 90 days until 200 days (7 months) of age (Allou et al., 2013).

Progesterone concentration

The results in Figure 2 to elucidate that plasma progesterone levels (ng/ml) as affected by FGA and PGF_{2α} (G2) and FGA, PGF_{2α}, PMSG and hCG (G3) as compared to the control group (G1). It could be observed that P4 levels attended to increase from the first day of pregnancy and rises to the end of the day before parturition. Also, progesterone was higher in G3 which contains the highest values of litter size at birth of newborn lambs as a result of increasing twinning rates in response to hormonal treatment for the superovulation. Mean concentration of P4 level was found to be 8.47 ng/ml in G3 followed by G2 (8.05 ng/ml) while the lowest value was observed in G1 (7.40 ng/ml).

Early pregnancy is a critical period for maternal recognition, and the lack of nutrients that stimulate an increase in circulating progesterone can increase early embryonic losses (Viñoles et al., 2012). According to Mann et al. (2006), most embryonic losses occur during the first few days after fertilization and during the implantation process, inadequate luteal function being one of the main causes. The maintenance of pregnancy in ruminants depends on the continued secretion of progesterone by the corpus luteum, which inhibits luteolysis. Progesterone deficiency due to primary luteal insufficiency has been reported as a cause of embryonic death (Mann and Lamming, 2001; Diskin and Morris, 2008). Increase concentrations of progesterone (P4) during meta-oestrus and early di-oestrus improves embryonic growth and the production of interferon- τ (IFN- τ) (Spencer, 2013 and 2016; Arosh et al., 2016), which in turn improves the relationship between embryo and uterus and increases embryonic survival rates (Mann et al., 2006).

The higher values of progesterone levels, found in G3 followed by G2 than that in the control group, may be due to the number of corpora lutea present as it is the source of progesterone in sheep. El-Tarabany (2012) found that progesterone concentration decreased significantly in ewes conceived single than in ewes conceived twin's fetuses by 34.3%. Also, Khan and Ludri (2002) found that in twin bearing goats' plasma progesterone level was significantly higher than in single bearing goats during all days of experiment. Manalu et al. (1998) reported that ewes with a high litter size had heavier placental tissue. Moreover, ewes with a high litter size had high ovulation rates that resulted in high number of corpora lutea as sources of progesterone during the embryonic phase of pregnancy and, probably, throughout the pregnancy period

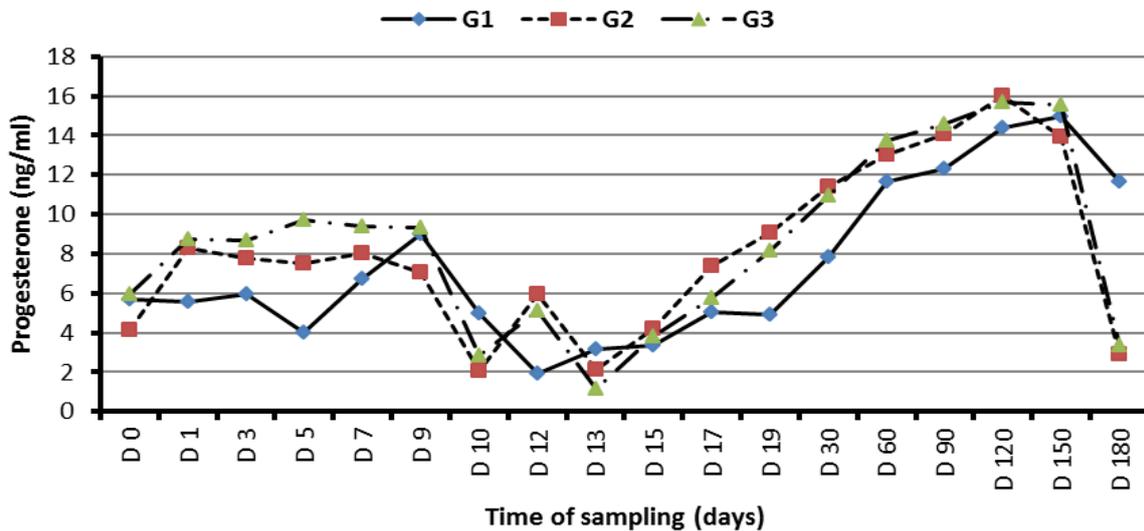


Figure 2. Plasma progesterone levels (ng/ml) as affected by FGA and PGF₂α (G2) and FGA, PGF₂α, PMSG and hCG (G3) compared to the control group (G1). G1 = control group; G2 = FGA and PGF₂α; G3 = FGA, PGF₂α, PMSG and hCG

Type of lambing and sex ratio

The data presented in **table 3** demonstrate that single rate was significantly higher in the control non treated group (G1) as compared to other treated groups, being 90.00, 86.96 and 64.00%, respectively. Results of present study indicated that twinning rate significantly increased in the hormonal manipulated group (G3) than (G2) and (G1) being 32.00, 13.04 and 10.00%, respectively, (**Table 3**). The influence of the superovulation treatment was observed for G3, as triple rate increased significantly in G3 as compared to other groups, with values of 4.0, 0.0 and 0.0%, respectively. Sex ratio (female %) exhibited significantly the highest values for G3 (60.00%) followed by G2 (59.09%) and finally G1 (46.15%). Conversely, sex ratio (male %) was significantly higher in G2 than in G1 and G3, recording 53.85, 40.91 and 40.00%, respectively.

A like to the Barki breed in our study herein, Awassi breed is a monotoccus with an ovulation rate of 1 and very low incidence of twinning (Jaber et al., 2004). For that reason, the twins and triplets born lambs weighed less at birth as compared with singles in control groups, this might be due to the competition between fetuses for nutrient supplied by the placenta from maternal circulation for growth due to the low capacity of uteri to provide for twins (Baneh and Hafezian, 2009). In Barki, twinning in general and especially triple births is not common. Moreover, Clement et al. (1997) in his study assumed that the longer of lambing interval is accompanied with the increase in the range of sizes with the rank of lambing. Herein this study triples birth in group 3 (G3) is probably accompanied with the effect of PMSG injection. Sheep male lambs' are known to be born heavier than females, (Gbangboché et al., 2005), who also reported that this trend in weight significant different may last up to 12 months or more, this observation is reported under same rearing conditions for birth weight for males and females, but these variations tends to disappear at 120 days of age (Poivey et al., 1982; Fall et al., 1983). Binns et al. (2002) and Brien et al. (2009) found higher mortality in male than female lambs. On the other hand, another study reported a higher survival risk in male lambs (Atashi et al., 2013), while mortality risks for gender were not statistically different in a study in Quebec (Arsenault et al., 2003).

Table 3. Types of lambing and sex ratio (%) of Barki ewes as affected by different hormonal protocols (LSM±SE)

Traits	Treatment (T)			Overall mean	± SEM	Significance
	G1	G2	G3			
Single rate	90.00 (18/20)	86.96 (20/23)	64.00 (16/25)	--	--	--
Twinning rate	10.00 ^b (2/20)	13.04 ^b (3/23)	32.00 ^a (8/25)	18.34	2.74	**
Triple rate	0 ^b	0 ^b	4.00 ^a (1/25)	--	--	--
Sex ratio (female)	59.09 (13/22)	46.15 (12/26)	60.00 (21/35)	55.42 (46/83)	1.62	*
Sex ratio (male)	40.91 (9/22)	53.85 (14/26)	40.00 (14/35)	44.58 (37/83)	1.62	*

G1 = control group; G2 = FGA and PGF₂α; G3 = FGA, PGF₂α, PMSG and hCG. Means within each row with different superscripts are significantly different at 5% level.

CONCLUSION

It could be concluded that, hormonal manipulation using intravaginal progestagen impregnated sponge and PMSG in the presence of hCG; for estrous synchronization and superovulation; would be a proper way for enhancing the reproductive efficiency of Barki ewes.

Acknowledgments

The authors are thankful to Dr. Khalid El-Bahrawy for his support and assisting in lab work and guide through writing the manuscript. Deepest thanks are due to Dr. Hamdy Gawish for facilitating the research work at the Sustainable Development Center for Matrouh Resources. We also would like to appreciate all participants who contributed during sample collection.

Competing interests

The authors declare that they have no conflict of interest with respect to the research, authorship, and/or publications of this article. The authors declare that they have no competing interests.

Author's contribution

Dr. Gamal Ashour designed the experiment, article writing and revision, Dr. Moharram Fouad El-Bassiony designed the experiment, laboratory analyses, statistical analysis, tabulation of experimental data, manuscript writing, commenting and approval, Dr. Sherif Mohamed Dessouki helped in statistical analysis, tabulation of experimental data and article writing; while, Mr. Mohamed Awad El-Wakeel helped in field study, collected data, laboratory analyses, manuscript writing. All authors have read and approved the final manuscript.

REFERENCES

- Abdul-Muin HB, Hasbudie B, Suraya MS, Panandam JM, Yaakub H, Theivanai J and Quaza Nizamuddin HN (2013). Effects of two CIDR-based oestrus synchronization protocols on oestrus response in Boer goats. *Malaysian Journal Animal Science*, 16(2): 29-35.
- Abecia JA, Forcada F and González-Bulnes A (2011). Pharmaceutical control of reproduction in sheep and goats. *Veterinary Clinics of North America: Food Animal Practice*, 27(1):67-79. DOI:10.1016/j.cvfa.2010.10.001
- Allou DS, Farougou S and Hountondji FCC (2013). Impact of prophylactic measures and the use of local food resources on the viability and growth of pre-weaning lambs in Djougou, in the northern region of Benin. *Animal and Plant Sciences*, 19(3): 2933-2940.
- Alvarado-Espino AS, Meza-Herrera CA, Carrillo E, González-Álvarez VH, Guillen-Muñoz JM, Ángel-García O and Mellado M (2016). Reproductive outcomes of Alpine goats primed with progesterone and treated with human chorionic gonadotropin during the anestrus-to-estrus transition season. *Animal Reproduction Science*, 167:133-138. DOI:10.1016/j.anireprosci.2016.02.019
- Andriyanto A and Manalu W (2011). Increasing goat productivity through the improvement of endogenous secretion of pregnant hormones using follicle stimulating hormone. *Animal Production*, 13(2): 89-93.
- Arosh JA, Banu SK and McCracken JA (2016). Novel concepts on the role of prostaglandins on luteal maintenance and maternal recognition and establishment of pregnancy in ruminants. *Journal of Dairy Science*, 99(7):5926-5940. DOI:10.3168/jds.2015-10335
- Arsenault J, Dubreuil P, Girard C, Simard C and Belanger D (2003). Maedi-visna impact on productivity in Quebec sheep flocks (Canada). *Preventive Veterinary Medicine*, 59: 125-137. Doi:https://doi.org/10.1016/S0167-5877(03)00086-2
- Atashi H, Izadifard J, Zamiri MJ and Akhlaghi A (2013). Investigation in early growth traits, litter size, and lamb survival in two Iranian fat-tailed sheep breeds. *Tropical Animal Health and Production*, 45: 1051-1054. DOI:10.1007/s11250-012-0308-9
- Baneh H and Hafezian H (2009). Effects of environmental factors on growth traits in Ghezel sheep. *African Journal of Biotechnology*, 8:2903-2907. Doi:10.5897/AJB09.284
- Binns SH, Cox IJ, Rizvi S and Green LE (2002). Risk factors for lamb mortality on UK sheep farms. *Preventive Veterinary Medicine*, 52: 287-303. Doi:https://doi.org/10.1016/S0167-5877(01)00255-0
- Brien FD, Hebart ML, Jaensch KS, Smith DH and Grimson RJ (2009). Genetics of lamb survival: a study of Merino resource flocks in South Australia. *Proceedings of Association for the Advancement of Animal Breeding and Genetics*, 18: 492-495. Doi:https://www.researchgate.net/profile/Forbes_Brien
- Carson AF, Irwin D and Kilpatrick DJ (2001). A comparison of Scottish Blackface and Cheviot ewes and five sire breeds in term of lamb output at weaning in hill sheep system. *The Journal of Agriculture Science*, 137: 221-233. Doi:https://doi.org/10.1017/S0021859601001277
- Chniter M, Hammadi M, Khorchani T, Krit R, Lahsoumi B, Sassi MB, Nowak R and Hamouda MB (2011). Phenotypic and seasonal factors influence birth weight, growth rate and lamb mortality in D'man sheep maintained under intensive management in Tunisian oases. *Small Ruminant Research*, 99:166-170. Doi:https://doi.org/10.1016/j.smallrumres.2011.03.046

- Chniter M, Hammadi M, Khorchani T, Sassi MB, Hamouda MB and Nowak R (2013). Aspects of neonatal physiology have an influence on lambs' early growth and survival in prolific D'man sheep. *Small Ruminant Research*, 111:162-170. Doi:<https://doi.org/10.1016/j.smallrumres.2012.10.004>
- Clément V, Poivey JP, Faugère O, Tillarde Lancelot R, Gueye A, Richard D and Bibe B (1997). Etude de la variabilité des caractères de reproduction chez les petits ruminants en milieu traditionnel au Sénégal. *Revue d'élevage et de médecine vétérinaire des pays tropicaux*, 50:235-249.
- Cognie Y, Baril G, Poulin N and Mermillod P (2003). Current status of embryo technologies in sheep and goat. *Theriogenology*. 59: 171-188. Doi:[https://doi.org/10.1016/S0093-691X\(02\)01270-0](https://doi.org/10.1016/S0093-691X(02)01270-0)
- Dendena M (2017). *Controlo Da Actividade Reprodutiva e Inseminação Artificial Em Ovelhas Da Raça Churra Galega Bragançana (Thesis)*. 53 p.
- Diskin MG and Morris DG (2008). Embryonic and early foetal losses in cattle and other ruminants. *Reproduction of Domestic Animals*, 43(2): 260–267. Doi:10.1111/j.1439-0531.2008.01171.x
- Duncan DB (1955). Multiple range and multiple F tests. *Biometrics*, 11: 1- 42.
- Elliott J, Sneddon J, Lee JA and Blache D (2011). Producers have a positive attitude toward improving lamb survival rates but may be influenced by enterprise factors and perceptions of control. *Livestock Science*, 140:103-110. Doi:<https://doi.org/10.1016/j.livsci.2011.02.015>
- El-Tarabany AA (2012). Physiological changes in ewes conceived single or twin's fetuses related with survivability of lambs. *Arab Journal of Nuclear Science and Applications*, 45(3): 1-12.
- Fall A, Diop M, Sandford J, Gueye E, Wissocq YJ, Durkin J and Trail JCM (1983). Etude sur la productivité de moutons Djallonké au Centre de Recherches Zootechniques de Kolda, au Sénégal: 2. Poids corporels, productivité des brebis et du troupeau. *Revue d'élevage et de médecine vétérinaire des pays tropicaux*, 36(3):283-289.
- Fonseca J, Bruschi J, Zambrini F, Demczuk E, Viana J and Palhão M (2005). Induction of synchronized estrus in dairy goats with different gonadotrophins. *Animal Reproduction*, 2: 50-53.
- Fornazari R, Mateus Ó, Correia T, Quintas H, Maurício R, Conradi A, Francisco L, Álvaro A and Valentim R (2018). Estrus synchronization and artificial insemination with fresh and chilled semen in Assaf ewes. *Agriculture Science*, 9:8-22. DOI:10.4236/as.2018.91002
- Gbangboche AB, Hornich JL, Adamou-N'Diaye M, Edoth AP, Fanir F, Abiola FA and Leroy PL (2005). Caractérisation et maîtrise des paramètres de la reproduction et de la croissance des ovins Djallonké (*Ovis aries*). *Annales de Médecine Vétérinaire*, 149:170-182.
- Gongnet GP (1996). Étude des causes de mortalité des agneaux Peul-Peul et Touabire avant sevrage dans la zone sylvo-pastorale au Sénégal. *Sciences Vétérinaires et Médecine Comparée*, 98:133-42.
- Hosseinipanah SM, Anvarian M, Mousavinia M, Alimardan M, Hamzei S and Zengir SBM (2014). Effects of progesterone in synchronization of estrus and fertility in Shal ewes in nonproductive season. *European Journal of Experimental Biology*, 4(1):83-86.
- Husein MQ, Ababneh MM and Haddad SG (2005). The effects of progesterone priming on reproductive performance of GnRH-PGF2 alpha-treated anestrus goats. *Reprod. Nutr. Dev.*, 45:689-698. DOI: 10.1051/rnd:2005053
- Jaber LS, Habre A and Rawda N (2004). The effect of water restriction on certain physiological parameters in Awassi sheep. *Small Ruminant Research*, 54:115-120. Doi:<https://doi.org/10.1016/j.smallrumres.2003.11.004>
- Jainudeen M, Wahid H and Hafez E (2000). Ovulation induction, embryo production and transfer. In: *Reproduction in Farm Animals*, 7th Edition. Lippincott Williams & Wilkins. Maryland. USA. 405-430.
- Kaya A, G-Nes E and Memili E (2018). Application of reproductive biotechnologies for sustainable production of livestock in Turkey. *Turkish Journal of Veterinary and Animal Sciences*, 42: 143-151. Doi:10.3906/vet-1706-66
- Kearl LC (1982). *Nutrient Requirements of Ruminants in Developing Countries*. Utah Agric. Exp. Sta. Utah State University, Logan, U S A.
- Khan JR and Ludri RS (2002). Hormonal profiles during periparturient period in single and twin fetus bearing goats. *Asian-Australian Journal of Animal Science*, 15(3):346-351. Doi:<https://doi.org/10.5713/ajas.2002.346>
- Koyuncu M and Alticekic SO (2010). Effects of progestagen and PMSG on estrous synchronization and fertility in Kivircik ewes during natural breeding season. *Asian-Australian Journal of Animal Science*, 23:308-311. Doi:<https://doi.org/10.5713/ajas.2010.90393>
- Kusina N, Tarwirei F, Hamudikuwanda H, Agumba G and Mukwena J (2000). A comparison of the effects of progesterone sponges and ear implants, PGF2alpha, and their combination on efficacy of estrus synchronization and fertility of Mashona goat does. *Theriogenology*, 53:1567-1580. Doi:[https://doi.org/10.1016/S0093-691X\(00\)00298-3](https://doi.org/10.1016/S0093-691X(00)00298-3)
- Letelier CA, Contreras-Solis I, Garcia-Fernandez RA, Sanchez MA, Garcia-Palencia P, Sanchez B, Ariznavarreta C, Tresguerres JAF, Flores JM and Gonzalez-Bulnes A (2011). Effects of oestrus induction with progestagens or prostaglandin analogues on ovarian and pituitary function in sheep. *Animal Reproduction Science*, 126:61-69. Doi:<https://doi.org/10.1016/j.anireprosci.2011.04.012>
- Manalu W, Sumaryadi MY, Sudjatmogo and Satyaningtjas AS (1998). Effect of superovulation on maternal serum progesterone concentration, uterine and fetal weights at weeks 7 and 15 of pregnancy in Javanese thin-tail ewes. *Small Ruminant Research*, 30:171–176. Doi:[https://doi.org/10.1016/S0921-4488\(98\)00121-7](https://doi.org/10.1016/S0921-4488(98)00121-7)
- Mann GE and Lamming GE (2001). Relationship between maternal endocrine environment, early embryo development and inhibition of the luteolytic mechanism in cows. *Reproduction*, 121(1):175–180. Doi:<https://www.ncbi.nlm.nih.gov/pubmed/11226041>
- Mann GE, Fray MD and Lamming GE (2006). Effects of time of progesterone supplementation on embryo development and interferon- τ production in the cow. *Veterinary Journal*, 171(3):500–503. DOI: 10.1016/j.tvjl.2004.12.005

- Mekuriaw Z, Assefa H, Tegegne A and Muluneh D (2016). Estrus response and fertility of Menz and crossbred ewes to single prostaglandin injection protocol. *Tropical Animal Health and Production*, 48:53-57. DOI:10.1007/s11250-015-0919-z
- Motlomelo K, Greyling J and Schwalbach L (2002). Synchronisation of oestrus in goats: the use of different progestagen treatments. *Small Ruminant Research*, 45:45-49. DOI:10.1016/S0921-4488(02)00113-X
- Navanukraw C, Khanthusaeng V, Kraison A and Uriyapongson S (2014). Estrous and ovulatory responses following cervical artificial insemination in Thainative goats given a new or once-used controlled internal drug release with human chorionic gonadotropin. *Tropical Animal Health and Production*, 46 (8):1441–1446. DOI:10.1007/s11250-014-0662-x
- Omotesse BO, Rekwot PI, Ate IU, Ayo JO, Kawu MU, Rwuaan JS, Nwannenna AI, Mustapha RA and Bello AA (2016). An update on oestrus synchronisation of goats in Nigeria. *Asian Pacific Journal of Reproduction*, 5: 96-101. Doi:<https://doi.org/10.1016/j.apjr.2016.01.002>
- Piwczynski D, Sitkowska B and Wisniewska E (2012). Application of classification trees and logistic regression to determine factors responsible for lamb mortality. *Small Ruminant Research*, 103:225-231. Doi:<https://doi.org/10.1016/j.smallrumres.2011.09.014>
- Poivey JP, Landais E and Berger Y (1982). Etude et amélioration génétique de la croissance des agneaux Djallonké Résultats obtenus au Centre de Recherches Zootechniques de Bouaké (Côte-d'Ivoire). *Revue d'élevage et médecine vétérinaire des pays tropicaux*, 35(4):421-433. Doi:<https://agritrop.cirad.fr/455195/1/ID455195.pdf>
- Riaz H, Sattar A, Arshad MA and Ahmad N (2012). Effect of synchronization protocols and GnRH treatment on the reproductive performance in goats. *Small Ruminant Research*, 104(1):151–155. Doi:<https://doi.org/10.1016/j.smallrumres.2011.10.008>
- SAS (2004). *Statistical analysis system User's Guide*. Release 9.1. SAS institute, Cary, North Carolina, USA.
- Souza A, Garcia-Guerra A and Adams G (2014). Advances in ovarian stimulation. In: *Proceedings of the 2014 AETA and CETA/ACTE Joint Convention*; Madison, WI, USA.
- Spencer TE (2013). Early pregnancy: concepts, challenges, and potential solutions. *Animal Frontiers*, 3(4):48–55.
- Spencer TE, Forde N and Lonergan P (2016). The role of progesterone and conceptus-derived factors in uterine biology during early pregnancy in ruminants. *Journal of Dairy Science*, 99(7):5941–5950. Doi:<https://doi.org/10.3168/jds.2015-10070>
- Valentim R, Rodrigues I, Montenegro T, Sacoto S, Azevedo J and Gomes MJ (2016). Artificial Insemination in Sheep and Goat. In: *Reproduction Management in Sheep and Goat*. Agrotec. Portugal. ISSN 2182-4401. 21:10-13.
- Viñoles C, Glover KMM, Paganoni BL, Milton JTB and Martin GB (2012). Embryo losses in sheep during short-term nutritional supplementation. *Reproduction and Fertility Development*, 24(8):1040–1047. Doi:<http://dx.doi.org/10.1071/RD11281>
- Waheeb RS, El-amrawi GA, Metwelly KK and El-sabbagh AM (2017). Synchronization of estrus in field conditions using progestagen sponge, GnRH, and PGF2 α in Barki ewes during breeding season. *Alexandria Veterinary Science*, 54(2): 1-4. DOI:10.5455/ajvs.273830
- Webb R, Garnsworthy P, Gong JG and Armstrong D (2004). Control of follicular growth: local interactions and nutritional influences. *Journal of Animal Science*, 82: E63-E74. DOI:10.2527/2004.8213_supplE63x
- Yavuzer Ü (2005). The possibilities of twice-yearly lambing of Awassi sheep ewes without using hormones in an organic animal production system. *Turkish Journal of Veterinary Animal Science*, 29:27-30.
- Zarkawi M (2001). Oestrous synchronisation and twinning rate of Syrian Awassi ewes treated with progestagen and PMSG during the breeding season. *New Zealand Journal of Agricultural Research*, 44: 159-163. Doi:<https://doi.org/10.1080/00288233.2001.9513472>
- Zohara BF, Azizunnesa, Islama F, Alam GS and Bari FY (2014). Comparison of estrus synchronization by PGF2 α and progestagen sponge with PMSG in indigenous ewes in Bangladesh. *GSTF International Journal of Veterinary Science*, 1 (1):27-37. Doi:<http://dl6.globalstf.org/index.php/JVet/article/view/1330/1248>

Instructions for Authors

Manuscript as Original Research Paper, Short Communication, Case Reports and Review or Mini-Review are invited for rapid peer-review publishing in the *World's Veterinary Journal* (WVJ). Considered subject areas include: Behavior; environment and welfare; animal reproduction and production; parasitology, endocrinology, microbiology, immunology, pathology, pharmacology, epidemiology, molecular biology, immunogenetics, surgery, radiology, ophthalmology, dermatology, chronic disease, anatomy, and non-surgical pathology issues of small to large animals, cardiology and oncology are sub-specialties of veterinary internal medicine. ... [view full aims and scope](#)

[WVJ EndNote Style](#)

[Manuscript Template \(.doc\)](#)

[Sample Articles](#)

[Declaration form](#)

[Policies and Publication Ethics](#)

Submission

The manuscript and other correspondence should be [submit online](#) preferentially. Please embed all figures and tables in the manuscript to become one single file for submission. Once submission is complete, the system will generate a manuscript ID and password sent to author's contact emails: editor@wvj.science-line.com. All manuscripts must be checked (by English native speaker) and submitted in English for evaluation (in totally confidential and impartial way).

Supplementary information:

The online submission form allows supplementary information to be submitted together with the main manuscript file and covering letter. If you have more than one supplementary files, you can submit the extra ones by email after the initial [submission](#). Author guidelines are specific for each journal. Our Word template can assist you by modifying your page layout, text formatting, headings, title page, image placement, and citations/references such that they agree with the guidelines of journal. If you believe your article is fully edited per journal style, please use our [Word template](#) before submission.

[Supplementary materials](#) may include figures, tables, methods, videos, and other materials. They are available online linked to the original published article. Supplementary tables and figures should be labeled with a "S", e.g. "Table S1" and "Figure S1". The maximum file size for supplementary materials is 10MB each. Please keep the files as small possible to avoid the frustrations experienced by readers with downloading large files.

Submission to the Journal is on the understanding that:

- 1.The article has not been previously published in any other form and is not under consideration for publication elsewhere;
- 2.All authors have approved the submission and have obtained permission for publish work.
- 3.Researchers have proper regard for conservation and animal welfare considerations (see [IAVE-Author Guidelines on Animal Ethics and Welfare](#)). Attention is drawn to the '[Guidelines for the Treatment of Animals in Research and Teaching](#)'. Any possible adverse consequences of the work for populations or individual organisms must be weighed against the possible gains in knowledge and its practical applications. If the approval of an ethics committee is required, please provide the name of the committee and the approval number obtained.

Ethics Committee Approval

Experimental research involving human or animals should have been approved by author's institutional review board or ethics committee. This information can be mentioned in the manuscript including the name of the board/committee that gave the approval. Investigations involving humans will have been performed in accordance with the principles of [Declaration of Helsinki](#). And the use of animals in experiments will have observed the Interdisciplinary Principles and Guidelines for the Use of Animals in Research, Testing, and Education by the New York Academy of Sciences, Ad Hoc Animal Research Committee. If the manuscript contains photos or parts of photos of patients, informed consent from each patient should be obtained. Patient's identities and privacy should be carefully protected in the manuscript.

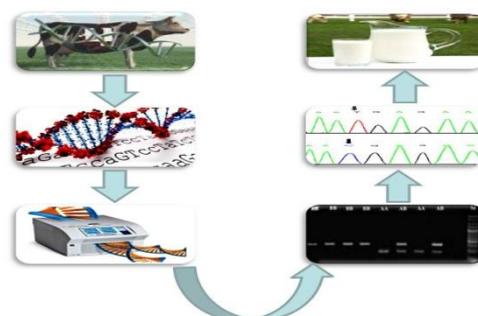
Competing Interests

Competing interests that might interfere with the objective presentation of the research findings contained in the manuscript should be declared in a paragraph heading "Competing interests" (after Acknowledgment section and before References). Examples of competing interests are ownership of stock in a company, commercial grants, board membership, etc. If there is no competing interest, please use the statement "The authors have declared that no competing interest exists."

Graphical Abstract:

Authors should provide a graphical abstract (a beautifully designed feature figure) to represent the paper aiming to catch the attention and interest of readers. Graphical abstract will be published online in the table of content. The graphical abstract should be colored, and kept within an area of 12 cm (width) x 6 cm (height) or with similar format. Image should have a minimum resolution of 300 dpi and line art 1200dpi. **Note:** Height of the image should be no more than the width. Please avoid putting too much information into the graphical abstract as it occupies only a small space. Authors can provide the graphical abstract in the format of PDF, Word, PowerPoint, jpg, or png, after a manuscript is accepted for publication. See more samples from [archive](#).

If you have decided to provide a Professional Graphical Abstract, please click here.



Presentation of the article

Main Format

First page of the manuscripts must be properly identified by the title and the name(s) of the author(s). It should be typed in Times New Roman (font sizes: 17pt in capitalization for the title, 10pt for the section headings in the body of the text and the main text, 9pt for References, double spaced, in A4 format with 2cm margins. All pages and lines of the main text should be numbered consecutively throughout the manuscript. The manuscript must be saved in a .doc format, (not .docx files). Abbreviations in the article title are not allowed.

Manuscripts should be arranged in the following order:

- a. TITLE (brief, attractive and targeted);
- b. Name(s) and Affiliation(s) of author(s) (including post code) and corresponding E-mail;
- c. ABSTRACT;
- d. Key words (separate by semicolons; or comma,);
- e. Abbreviations (used in the manuscript);
- f. INTRODUCTION;
- g. MATERIALS AND METHODS;
- h. RESULTS;
- i. DISCUSSION;
- j. CONCLUSION;
- k. Acknowledgements (if there are any);
- l. REFERENCES;
- m. Tables;
- n. Figure captions;
- o. Figures;

Results and Discussion can be presented jointly if preferred.

Discussion and Conclusion can be presented jointly if preferred.

Article Sections Format

Title should be a brief phrase describing the contents of the paper. The first letter of each word in title should use upper case. The Title Page should include the author(s)'s full names and affiliations, the name of the corresponding author along with phone and e-mail information. Present address (es) of author(s) should appear as a footnote.

Abstract should be informative and completely self-explanatory, briefly present the topic, state the scope of the experiments, indicate significant data, and point out major findings and conclusions. The abstract should be 150 to 300 words in length. Complete sentences, active verbs, and the third person should be used, and the abstract should be written in the past tense. Standard nomenclature should be used and abbreviations should be avoided. No literature should be cited. Following the abstract, about 3 to 10 **key words** that will provide indexing references should be listed.

Introduction should provide a clear statement of the problem, the relevant literature on the subject, and the proposed approach or solution. It should be understandable to colleagues from a broad range of scientific disciplines.

Materials and Methods should be complete enough to allow experiments to be reproduced. However, only truly new procedures should be described in detail; previously published procedures should be cited, and important modifications of published procedures should be mentioned briefly. Capitalize trade names and include the manufacturer's name and address. Subheadings should be used. Methods in general use need not be described in detail.

Results should be presented with clarity and precision. The results should be written in the past tense when describing findings in the author(s)'s experiments. Previously published findings should be written in the present tense. Results should be explained, but largely without referring to the literature. Discussion, speculation and detailed interpretation of data should not be included in the results but should be put into the discussion section.

Discussion should interpret the findings in view of the results obtained in this and in past studies on this topic. State the conclusions in a few sentences at the end of the paper. The Results and Discussion sections can include subheadings, and when appropriate, both sections can be combined.

Conclusion can be presented jointly if preferred.

Acknowledgments of persons, grants, funds, etc should be brief.

Tables should be kept to a minimum and be designed to be as simple as possible. Tables are to be typed double-spaced throughout, including headings and footnotes. Each table should be on a separate page, numbered consecutively in Arabic numerals and supplied with a heading and a legend. Tables should be self-explanatory without reference to the text. The details of the methods used in the experiments should preferably be described in the legend instead of in the text. The same data should not be presented in both table and graph forms or repeated in the text.

Figure legends should be typed in numerical order on a separate sheet. Graphics should be prepared using applications capable of generating high resolution GIF, TIFF, JPEG or PowerPoint before pasting in the Microsoft Word manuscript file. Use Arabic numerals to designate figures and upper case letters for their parts (Figure 1). Begin each legend with a title and include sufficient description so that the figure is understandable without reading the text of the manuscript. Information given in legends should not be repeated in the text.

Declarations section - Please include declarations heading

Please ensure that the sections:

-Ethics (and consent to participate)

-Authors' contributions

-Competing interests

-Availability of data and materials

are included at the end of your manuscript in a Declarations section.

Authors' Contributions

For manuscripts with more than one author, WVJ require an Authors' Contributions section to be placed after the Competing Interests section.

An 'author' is generally considered to be someone who has made substantive intellectual contributions to a published study. To qualify as an author one should 1) have made substantial contributions to conception and design, or acquisition of data, or analysis and interpretation of data; 2) have been involved in drafting the manuscript or revising it critically for important intellectual content; and 3) have given final approval of the version to be published. Each author should have participated sufficiently in the work to take public responsibility for appropriate portions of the content. Acquisition of funding, collection of data, or general supervision of the research group, alone, does not justify authorship.

We suggest the following format (please use initials to refer to each author's contribution): AB carried out the molecular genetic studies, participated in the sequence alignment and drafted the manuscript. JY carried out the immunoassays. MT participated in the sequence alignment. ES participated in the design of the study and performed the statistical analysis. FG conceived of the study, and participated in its design and coordination and helped to draft the manuscript. All authors read and approved the final manuscript.

For authors that equally participated in a study please write '[All/Both authors contributed equally to this work.](#)' Contributors who do not meet the criteria for authorship should be listed in an acknowledgements section.

Competing Interests

Competing interests that might interfere with the objective presentation of the research findings contained in the manuscript should be declared in a paragraph heading "Competing interests" (after Acknowledgment section and before References). Examples of competing interests are ownership of stock in a company, commercial grants, board membership, etc. If there is no competing interest, please use the statement "[The authors declare that they have no competing interests.](#)".

World's Veterinary Journal adheres to the definition of authorship set up by The International Committee of Medical Journal Editors (ICMJE). According to the ICMJE authorship criteria should be based on 1) substantial contributions to conception and design of, or acquisition of data or analysis and interpretation of data, 2) drafting the article or revising it critically for important intellectual content and 3) final approval of the version to be published. Authors should meet conditions 1, 2 and 3.

It is a requirement that all authors have been accredited as appropriate upon submission of the manuscript. Contributors who do not qualify as authors should be mentioned under Acknowledgements.

Change in authorship

We do not allow any change in authorship after provisional acceptance. We cannot allow any addition, deletion or change in sequence of author name. We have this policy to prevent the fraud.

Acknowledgements

We strongly encourage you to include an Acknowledgements section between the Authors' contributions section and Reference list. Please acknowledge anyone who contributed towards the study by making substantial contributions to conception, design, acquisition of data, or analysis and interpretation of data, or who was involved in drafting the manuscript or revising it critically for important intellectual content, but who does not meet the criteria for authorship. Please also include their source(s) of funding. Please also acknowledge anyone who contributed materials essential for the study.

Authors should obtain permission to acknowledge from all those mentioned in the Acknowledgements. Please list the source(s) of funding for the study, for each author, and for the manuscript preparation in the acknowledgements section. Authors must describe the role of the funding body, if any, in study design; in the collection, analysis, and interpretation of data; in the writing of the manuscript; and in the decision to submit the manuscript for publication.

Data Deposition

Nucleic acid sequences, protein sequences, and atomic coordinates should be deposited in an appropriate database in time for the accession number to be included in the published article. In computational studies where the sequence information is unacceptable for inclusion in databases because of lack of experimental validation, the sequences must be published as an additional file with the article.

References

1. A WVJ reference style for [EndNote](#) may be found [here](#).
2. All references to publications made in the text should be presented in a list with their full bibliographical description.
3. In the text, a reference identified by means of an author's name should be followed by the date of the reference in parentheses. When there are more than two authors, only the first author's surname should be mentioned, followed by 'et al'. In the event that an author cited has had two or more works published during the same year, the reference, both in the text and in the reference list, should be identified by a lower case letter like 'a' and 'b' after the date to distinguish the works.
4. References in the text should be arranged chronologically (e.g. Kelebeni, 1983; Usman and Smith, 1992 and Agindotan et al., 2003). The list of references should be arranged alphabetically on author's surnames, and chronologically per author. If an author's name in the list is also mentioned with co-authors, the following order should be used: Publications of the single author, arranged according to publication dates - publications of the same author with one co-author - publications of the author with more than one co-author. Publications by the same author(s) in the same year should be listed as 1992a, 1992b, etc.
5. Names of authors and title of journals, published in non-latin alphabets should be transliterated in English.

6. A sample of standard reference is " 1th Author surname A, 2th Author surname B , 3th Author surname C. 2013. Article title should be regular and 7 pt . *World Vet. J.*, Add No. of Volume (Issue No.): 00-00."

7. The color of [references in the text](#) of article is [dark blue](#). Example: ([Preziosi et al., 2002](#); [Mills et al., 2015](#)).

-Examples (at the text)

Abayomi (2000), Agindotan et al. (2003), (Kelebeni, 1983), (Usman and Smith, 1992), (Chege, 1998; Chukwura, 1987a,b; Tijani, 1993,1995), (Kumasi et al., 2001).

--Examples (at References section)

a) For journal:

Lucy MC (2000). Regulation of ovarian follicular growth by somatotropin and insulin- like growth factors in cattle. *Journal of Dairy Science*, 83: 1635-1647.

Kareem SK (2001). Response of albino rats to dietary level of mango cake. *J. Agric. Res. Dev.* pp 31-38.

Chikere CB, Omoni VT and Chikere BO (2008). Distribution of potential nosocomial pathogens in a hospital environment. *African Journal of Biotechnology*. 7: 3535-3539.

b) For symposia reports and abstracts:

Cruz EM, Almatar S, Aludul EK and Al-Yaqout A (2000). Preliminary Studies on the Performance and Feeding Behaviour of Silver Pomfret (*Pampus argentens euphrasen*) Fingerlings fed with Commercial Feed and Reared in Fibreglass Tanks. *Asian Fisheries Society Manila, Philippine* 13: 191-199.

c) For edited symposia, special issues, etc., published in a journal:

Korevaar H (1992). The nitrogen balance on intensive Dutch dairy farms: a review. In: A. A. Jongebreur et al. (Editors), *Effects of Cattle and Pig Production Systems on the Environment: Livestock Production Science*, 31: 17-27.

d) For books:

AOAC (1990). Association of Official Analytical Chemists. *Official Methods of Analysis*, 15th Edition. Washington D.C. pp. 69-88.

Pelczar JR, Harley JP, Klein DA (1993). *Microbiology: Concepts and Applications*. McGraw-Hill Inc., New York, pp. 591-603.

e) Books, containing sections written by different authors:

Kunev M (1979). Pig Fattening. In: A. Alexiev (Editor), *Farm Animal Feeding*. Vol. III. Feeding of Different Animal Species, Zemizdat, Sofia, p. 233-243 (Bg).

In referring to a personal communication the two words are followed by the year, e.g. (Brown, J. M., personal communication, 1982). In this case initials are given in the text.

Nomenclature and Abbreviations

Nomenclature should follow that given in NCBI web page and Chemical Abstracts. Standard abbreviations are preferable. If a new abbreviation is used, it should be defined at its first usage. Abbreviations should be presented in one paragraph, in the format: "term: definition". Please separate the items by ";".

E.g. ANN: artificial neural network; CFS: closed form solution...

Abbreviations of units should conform to those shown below:

Decilitre	dl	Kilogram	kg
Milligram	mg	hours	h
Micrometer	mm	Minutes	min
Molar	mol/L	Mililitre	ml
Percent	%		

Other abbreviations and symbols should follow the recommendations on units, symbols and abbreviations: in "A guide for Biological and Medical Editors and Authors (The Royal Society of Medicine London 1977).

Papers that have not been published should be cited as "unpublished". Papers that have been accepted for publication, but not yet specified for an issue should be cited as "to be published". Papers that have been submitted for publication should be cited as "submitted for publication".

Formulae, numbers and symbols

1. Typewritten formulae are preferred. Subscripts and superscripts are important. Check disparities between zero (0) and the letter O, and between one (1) and the letter I.
2. Describe all symbols immediately after the equation in which they are first used.
3. For simple fractions, use the solidus (/), e.g. 10 /38.
4. Equations should be presented into parentheses on the right-hand side, in tandem.
5. Levels of statistical significance which can be used without further explanations are *P < 0.05, **P < 0.01, and ***P < 0.001
6. In the English articles, a decimal point should be used instead of a decimal comma.
7. In chemical formulae, valence of ions should be given, e.g. Ca²⁺ and CO₃²⁻, not as Ca⁺⁺ or CO₃.
8. Numbers up to 10 should be written in the text by words. Numbers above 1000 are recommended to be given as 10 powered x.
9. Greek letters should be explained in the margins with their names as follows: Αα - alpha, Ββ - beta, Γγ - gamma, Δδ - delta, Εε - epsilon, Ζζ - zeta, Ηη - eta, Θθ - theta, Ιι - iota, Κκ - kappa, Λλ - lambda, Μμ - mu, Νν - nu, Ξξ - xi, Οο - omicron, Ππ - pi, Ρρ - rho, Σσ - sigma, Ττ - tau, Υυ - ipsilon, Φφ - phi, Χχ - chi, Ψψ - psi, Ωω - omega.

Review/Decisions/Processing

Firstly, all manuscripts will be checked by [Docol@c](#), a plagiarism finding tool. A single blind reviewing model is used by WVJ for non-plagiarized papers. The manuscript is edited and reviewed by the English language editor and three reviewers selected by section editor of WVJ respectively. Also, a reviewer result form is filled by reviewer to guide authors. Possible decisions are: accept as is, minor revision, major revision, or reject. See sample of [evaluation form](#). Authors should submit back their revisions within 14 days in the case of minor revision, or 30 days in the case of major revision.

To submit a revision please [sign in here](#), fill out the form, and mark " Revised" attach the revision (MS word) and submit when completed.

After review and editing the article, a final formatted proof is sent to the corresponding author once again to apply all suggested corrections during the article process. The editor who received the final revisions from the corresponding authors shall not be hold responsible for any mistakes shown in the final publication. Manuscripts with significant results are typically reviewed and published at the highest priority.

Plagiarism: There is a zero-tolerance policy towards plagiarism (including self-plagiarism) in our journals. Manuscripts are screened for plagiarism by [Docol@c](#) a plagiarism finding tool, before or during publication, and if found they will be rejected at any stage of processing. See sample of [Docol@c-Report](#).

Declaration

After manuscript accepted for publication, a [declaration form](#) will be sent to the corresponding author who that is responsible to coauthors' agreements to publication of submitted work in WVJ after any amendments arising from the peer review.

Date of issue

The journal will be issued on 25th of March, June, September and December, each year.

Publication charges

No peer-reviewing charges are required. However, there is a \$95 editor fee for the processing of each primary accepted paper. Payment can be made by credit card, bank transfer, money order or check. Instruction for payment is sent during publication process as soon as manuscript is accepted.

The submission fee will be waived for invited authors, authors of hot papers, and corresponding authors who are editorial board members of the *World's Veterinary Journal* (WVJ). The Journal will consider requests to waive the fee for cases of financial hardship (for high quality manuscripts and upon acceptance for publication). Requests for waiver of the submission fee must be submitted via individual cover letter by the corresponding author and cosigned by an appropriate institutional official to verify that no institutional or grant funds are available for the payment of the fee. Letters including the manuscript title and manuscript ID number should be sent to: editor.wvj@gmail.com. It is expected that waiver requests will be processed and authors will be notified within one business day.

Submission Preparation Checklist

- Authors are required to check off their submission's compliance with all of the following items, and submissions may be returned to authors that do not adhere to the following guidelines.
- The submission has not been previously published, nor is it before another journal for consideration (or an explanation has been provided in Comments to the Editor).
- The submission file is in Microsoft Word, RTF, or PDF document file format.
- Where available, URLs for the references have been provided.
- The text is single-spaced; uses a 12-point font; and all illustrations, figures, and tables are placed within the text at the appropriate points, rather than at the end.
- The text adheres to the stylistic and bibliographic requirements outlined in the Author Guidelines.

SCIENCELINE PUBLISHING CORPORATION

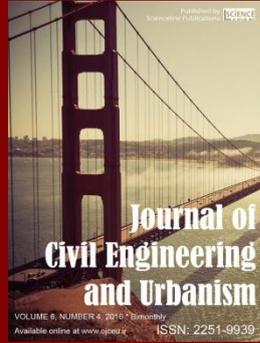
Scienceline Publication Ltd is a limited liability non-profit non-stock corporation incorporated in Turkey, and also is registered in Iran. Scienceline journals that concurrently belong to many societies, universities and research institutes, publishes internationally peer-reviewed open access articles and believe in sharing of new scientific knowledge and vital research in the fields of life and natural sciences, animal sciences, engineering, art, linguistic, management, social and economic sciences all over the world. Scienceline journals include:

Online Journal of Animal and Feed Research



ISSN 2228-7701; Bi-monthly
[View Journal](#) | [Editorial Board](#)
 Email: editors@ojaf.r.ir
[Submit Online >>](#)

Journal of Civil Engineering and Urbanism



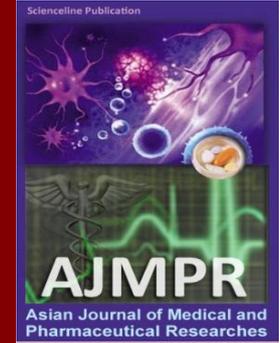
ISSN 2252-0430; Bi-monthly
[View Journal](#) | [Editorial Board](#)
 Email: ojceu@ojceu.ir
[Submit Online >>](#)

Journal of Life Sciences and Biomedicine



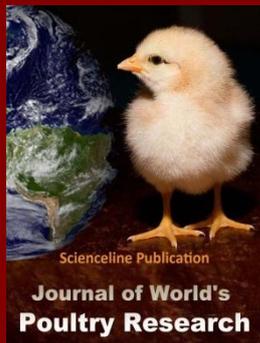
ISSN: 2251-9939; Bi-monthly
[View Journal](#) | [Editorial Board](#)
 Email: editors@jlsb.science-line.com
[Submit Online >>](#)

Asian Journal of Medical and Pharmaceutical Researches



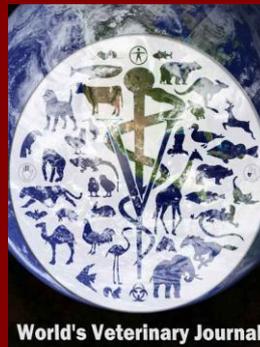
ISSN: 2322-4789; Quarterly
[View Journal](#) | [Editorial Board](#)
 Email: editor@ajmpr.science-line.com
[Submit Online >>](#)

Journal of World's Poultry Research



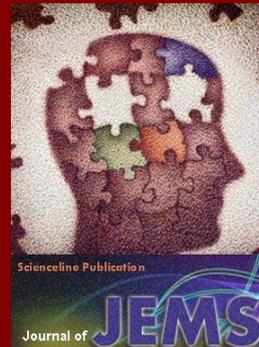
ISSN: 2322-455X; Quarterly
[View Journal](#) | [Editorial Board](#)
 Email: editor@jwpr.science-line.com
[Submit Online >>](#)

World's Veterinary Journal



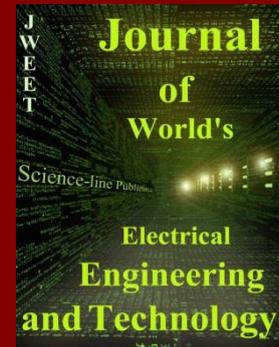
ISSN: 2322-4568; Quarterly
[View Journal](#) | [Editorial Board](#)
 Email: editor@wjv.science-line.com
[Submit Online >>](#)

Journal of Educational and Management Studies



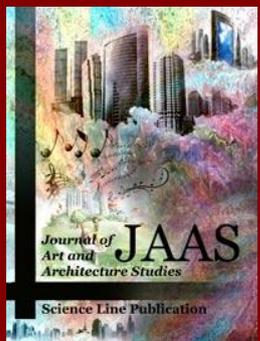
ISSN: 2322-4770; Quarterly
[View Journal](#) | [Editorial Board](#)
 Email: info@jems.science-line.com
[Submit Online >>](#)

Journal of World's Electrical Engineering and Technology



ISSN: 2322-5114; Irregular
[View Journal](#) | [Editorial Board](#)
 Email: editor@jweet.science-line.com
[Submit Online >>](#)

Journal of Art and Architecture Studies



ISSN: 2383-1553; Irregular
[View Journal](#) | [Editorial Board](#)
 Email: jaas@science-line.com
[Submit Online >>](#)

Asian Journal of Social and Economic Sciences



ISSN: 2383-0948; Quarterly
[View Journal](#) | [Editorial Board](#)
 Email: ajses@science-line.com
[Submit Online >>](#)

Journal of Applied Business and Finance Researches



ISSN: 2382-9907; Quarterly
[View Journal](#) | [Editorial Board](#)
 Email: jabfr@science-line.com
[Submit Online >>](#)

Scientific Journal of Mechanical and Industrial Engineering



ISSN: 2383-0980; Quarterly
[View Journal](#) | [Editorial Board](#)
 Email: sjmie@science-line.com
[Submit Online >>](#)